

## Simple and sensitive high performance liquid chromatographic method for the simultaneous quantitation of the phenylalanine in human plasma

Hossein Danafar<sup>1,2\*</sup>, Mehrdad Hamidi<sup>3</sup>

<sup>1</sup> Zanzan Pharmaceutical Nanotechnology Research Center, Zanzan University of Medical Sciences, Zanzan, Iran

<sup>2</sup> Department of Medicinal Chemistry, School of Pharmacy, Zanzan University of Medical Sciences, Zanzan, Iran

<sup>3</sup> Department of Pharmaceutics, School of Pharmacy, Zanzan University of Medical Sciences, Zanzan, Iran

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### Abstract

Phenylalanine (Phe) is the most reliable indicator for the diagnosis of phenylketonuria (PKU). The purpose of this study is to establish a reliable and quick method for the assignment of Phe in peripheral capillary blood from newborns and children by high performance liquid chromatography with ultraviolet detection (HPLC-UV). PKU is an inborn error of metabolism characterized by the inability of the body to use Phe. A rapid and sensitive high performance liquid chromatographic (HPLC) method has been developed for determination of Phe in plasma. The method uses a protein precipitation step with sulfosalicylic acid for sample preparation by separation on a Nova-pack C<sub>18</sub> column using sodium acetate buffer and acetonitrile (94: 6 v/v) adjusted to pH 6.5 with glacial acetic acid. The eluted peaks detected by a UV detector was set at wavelength of 215 nm. The method was validated in the range of Phe concentrations from 0.1 to 20 µg/ml. The limits of detection (LOD) and quantitation (LOQ) of the method were 0.05 and 0.1 µg/ml, respectively. The average drug recovery from plasma was 88.60 percent throughout the linear concentration range., with the average within-run and between-run accuracy values of 103.3 and 115.350, respectively. The method is quick, easy, very steady and precise for the screen, assignment, and evaluation of Phe in human plasma by HPLC, which is particularly a useful way for screening and diagnosis of PKU and monitoring of a diet therapy.

**Keywords:** Phenylketonuria (PKU); phenylalanine (phe) assay, reversed-phase HPLC

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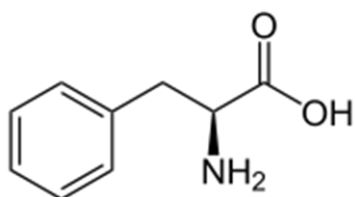
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### Introduction

Phenylketonuria (PKU) is an inborn error of metabolism characterized by the inability of the body to use phenylalanine (Phe) (Fig. 1). The inability to hydrolyze Phe to tyrosine (Tyr) may adversely affect the synthesis of tyrosine dependent neurotransmitter substances (1). Left untreated, severe mental retardation result (2). PKU is investigated by studied levels of Phe in the blood. The range levels were observed in classic PKU from 6 to 80

mg/dl in humans (3). PKU shows a spectrum of recessively inherited metabolic disorders where the transformation of the aromatic amino acid Phe to Tyr, which is the precursor of catecholamines, is phenylalanineired (4,5). Phe is known as essential aromatic acids in mammals. Another aromatic amino acid, Tyr is not important when sufficient Phe is available, because the amino acid is normally

\* E-mail: [danafar@zums.ac.ir](mailto:danafar@zums.ac.ir)



**Figure 1** Chemical structure of phenylalanine

synthesized from Phe in the liver. Measurement of Phe levels in blood is an important way for the diagnosis and subsequent dietary management of PKU. Moreover simultaneous detection of blood Phe and Tyr concentrations and calculating the ratio of Phe/Tyr can reduce the false positive rate in PKU screening (6,7). Currently, high-performance liquid-chromatographic (HPLC) assay still roles an extremely important role in the assignment of Phe, and a variety of HPLC methods are used to the simultaneous determination of Phe in serum, plasma and dried blood-spot specimens (8-20)The evaluating and determination of PKU need to the measure of blood Phe levels using HPLC methods have been used for this measurement; however, part of these methods have shortages including instability of samples, cost of agents and rigidity encountered in sample preparation (21-26). In contrast, the HPLC methods used in this study is cheap, sensitive, quick, accurate and available. The objective of this study is to determine the blood Phe level in PKU humans using HPLC.

## Materials and Methods

### Materials

5-Sulfosalicylic acid (SSA), sodium acetate, glacial acetic acid, L-phenylalanine and HPLC-grade acetonitrile and methanol were obtained from Merck (Darmstadt, Germany). Other chemicals

and solvents were from chemical lab or HPLC purity grades, whenever needed, and were purchased locally.

### Instrument and HPLC method

The HPLC system to include of pump (KNAUER, model 1000, Germany), wavelength UV detector (KNAUER, model 2800 (DAD), Germany) used at a wavelength of 215 nm with the outputs to record and analyze using with a software (ChromGate, KNAUER, Germany). The drug analization was performed using a C<sub>18</sub>analytical column (250mm×4.6mm, particle size 5µm; Perfectsill, MZ-Analysentechnik, Germany) equipped by a guard column of the same packing. The mobile phase sodium acetate buffer and acetonitrile (94: 6 v/v) adjusted to pH 6.5 with glacial acetic acid was with a flow rate of 1 ml/min. Sample injection to system (20 µL) was made by a loop.

### Standard preparation

A stock solution of 10 µg/ml Phe in water was prepared, from which the concentrations of 20, 10, 5, 2.5, 1, 0.5 and 0.1 µg/ml were prepared by serially diluting this solution with the amount of water. A series of plasma samples with Phe concentrations of 20, 10, 5, 2.5, 1, 0.5 and 0.1 µg/ml were prepared by 1:10 dilution of the commended solutions with drug-free human plasma.

### Assay procedure

To 500 µL plasma samples were added, to 400 µL water and 100 µL 30% sulfosalicylic acid. The mixture was vortexed for 30 s. After centrifuged at 12000 rpm for 15 min, the supernatant was isolated and a sample of 20 µL was injected into the HPLC immediately.

### Analysis Validation Tests

a) *Standard curve (Linear range)*

The plasma samples with a series of known concentrations, prepared as commended, were analyzed in three runs and, in each case, the linear regression analysis was achieved on known concentrations of Phe against the corresponding peak heights and, then, the regression coefficient ( $r$ ), slope, and  $y$ -intercept of the resulting calibration curves were calculated.

*b) Within-run variations*

In one run, three samples with concentrations of 10, 2.5, and 0.5  $\mu\text{g/ml}$  (from high, middle, and low regions of the standard curve) were prepared in triplicate and analyzed by HPLC method. Then, the coefficient of variations (CV %) of the corresponding determined concentrations were determined in each case.

*c) Between-run variations*

On three different runs, samples from upper, intermediate, and lower concentration regions used for construction of standard curve (the same as within-run variations test) were prepared and analyzed by HPLC method. Then, the corresponding CV% values were determined.

*d) Absolute recovery (accuracy)*

For each sample tested for within- and between-run variations, the absolute recovery of the method was calculated as the percent ratio of the measured concentration (determined using standard curve) to the corresponding nominal added concentration.

*e) Relative recovery (matrix effect)*

Three samples with concentrations of 10, 2.5, and 0.5  $\mu\text{g/ml}$  (from high, middle, and low regions of the standard curve) were prepared in triplicate and analyzed by HPLC method. Then, the ratio of the recorded peak heights to the peak heights calculated from the direct injection of the aqueous solutions of Phe with the same concentrations were determined as percentage in each case.

*f) Limits of detection and quantitation*

Limit of detection (LOD) was calculated at the lowest concentration to be detected, taking into investigation of a signal-to-baseline noise ratio of 3. Limit of quantitation (LOQ) was applied as the lowest concentration at which the precision to state by relative standard deviation (R.S.D.) is lower than 20% and accuracy to state by relative difference of the measured and true value (RE) is also lower than 20%. The LOQ was repeated five times for confirmation.

## **Results**

### ***Method development***

To study of the complex biological matrix of the samples to be analyzed and the nature of the method to be applied for drug assay, the method development efforts were made in two different areas of sample preparation and analyte separation which are studied in detail in the following sections:

*a) Sample preparation*

Protein precipitation was need and important because this technique can not only purify but also concentrate the sample. sulfosalicylic acid, percholoric acid and water were all exanimate and water: sulfosalicylic acid (80:20 v/v) was finally selected because of its high extraction efficiency and less interference.

*b) Analyte separation*

In response to lack of an available, sensitive, and easy-to-use analysis method for Phe assay in plasma as an essential part of pharmacokinetic and bioequivalence evaluation projects on the drug we selected a simple and reliable HPLC method with UV detection based on the available equipments found in most pharmaceutical laboratories. Finally, initially a series of isocratic as well as gradient conditions using different usual mobile phase compositions, polarities, ionic strengths,

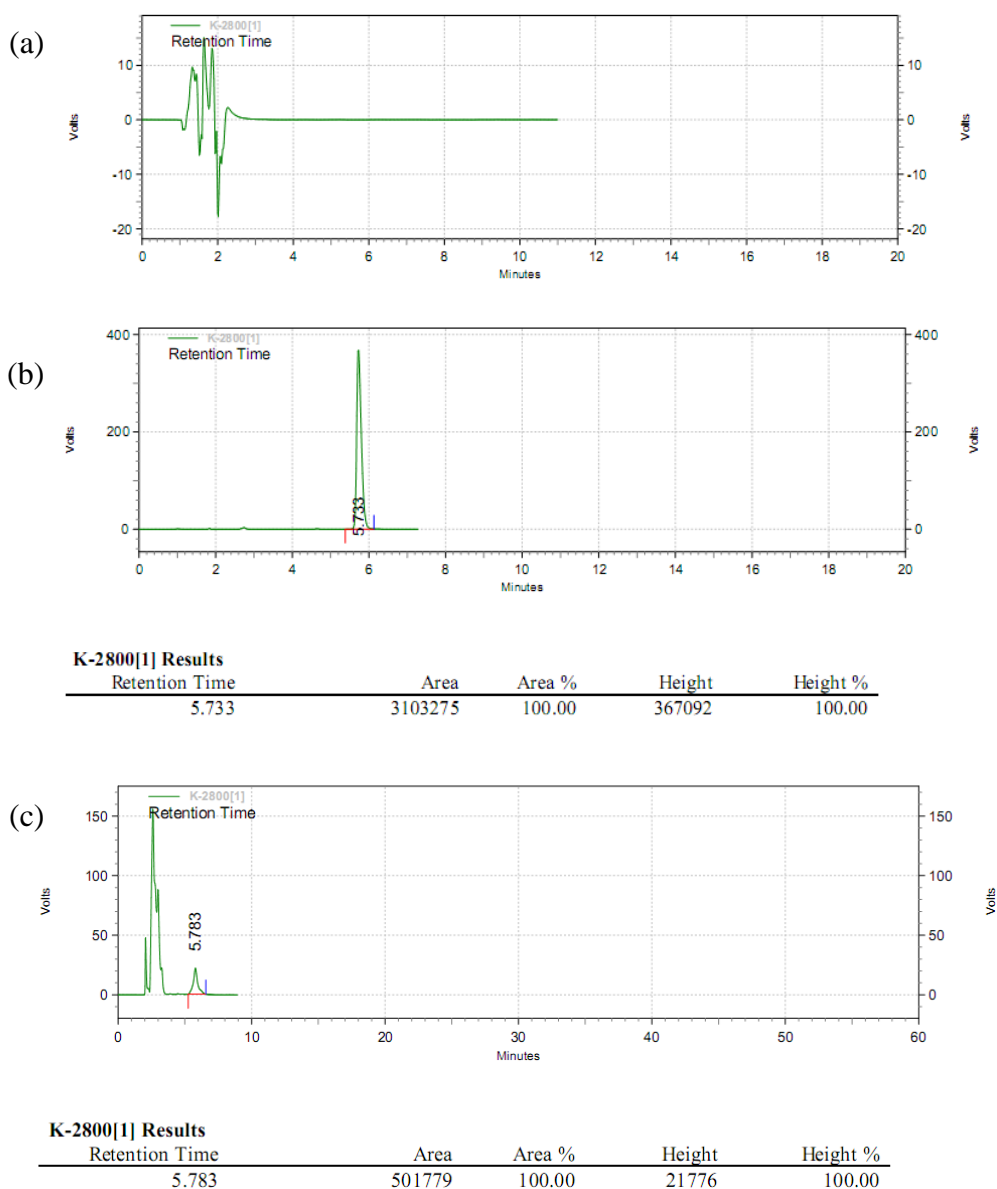
and pH values were examined in order to determine the best condition for the analyte separation. Typical chromatograms produced from the developed method are shown in Fig. 2.

**Method validation tests**

*a) Linearity*

The method produced linear responses

throughout the Phe concentration range of 0.1-20 µg/ml, which is suitable for intended purposes. A typical linear regression equation of the method was:  $y = 0.0215x + 0.194$ , with x and y representing Phe concentration (in µg/ml) and peak height (in arbitrary units), respectively, and the regression coefficient ( $r^2$ ) of 0.9915 (Fig. 3).



**Figure 2** Typical chromatograms of the HPLC method developed for phenylalanine assay in human plasma: a) Chromatogram of stock solution b) human plasma spiked to a 5µg/ml drug concentration; C) human blank plasma

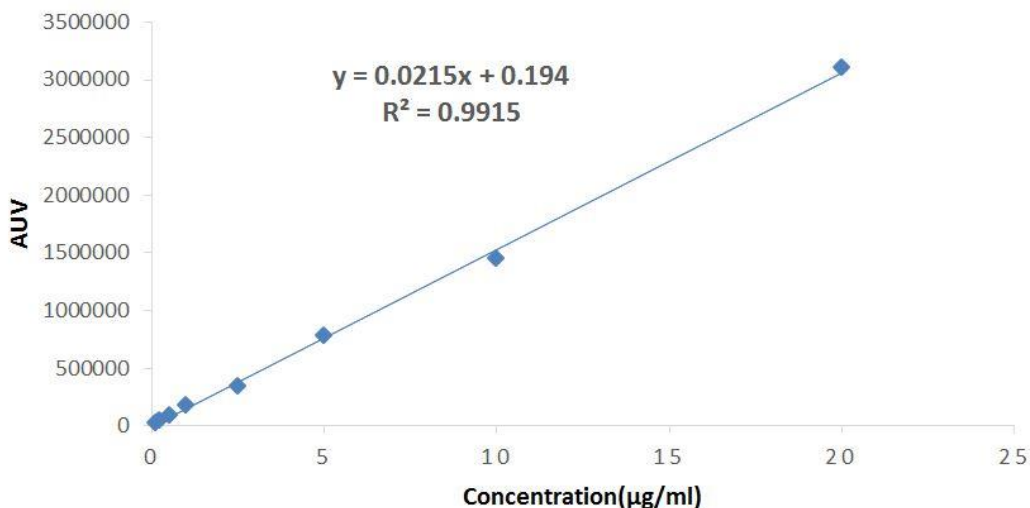


Figure 3 Standard curve of phenylalanine

*b) Within-run variations and accuracy*

The within-run variations of the HPLC method as well as the corresponding absolute recoveries are shown in Table 1.

*c) Between-run variations and accuracy*

The between-run variations of the HPLC method as well as the corresponding absolute recoveries are shown in Table 2.

*d) Relative recovery*

The relative recovery of Phe using the assay method is shown in Table 3.

*e) Limit tests*

The limits of detection (LOD) and quantization (LOQ) of the method were 0.05 and 0.1 µg/ml, respectively.

In general, the results of the validation tests showed that the method has a high degree of accuracy, repeatability, reproducibility, and recovery with application limits being in the desired range for routine applications.

**Discussion**

Untreated PKU patients may lead to mental retardation, microcap and other serious medical problems. However, if

PKU patient is degassed early enough; they can grow up with normal brain development by treatment and monitoring Phe levels through mixture of diet and medication. Detection of Phe concentration in blood is the chosen method for diagnosis of PKU and treatment assignment. Methods for quantitative assignment of Phe in blood samples include HPLC method (8–20), fluorometric method (27), enzymatic method (28–30). Among these methods, HPLC is currently regarded as a simple, accurate, and rapid method to screen amino acid metabolic disorders in newborn. However, these methods declared above have some disadvantages, containing requirements of complex reprivatizing agents or sophisticated sample preparation (12-15). For example, phenyl isothiocyanate reagent was applied for the pre column derivatization of the amino acids (16). For the diagnosing of PKU or monitoring of in PKU patients, repeated Phe determinations are required. To achieve this goal, it is highly desired to have a single, simple and inexpensive analytical method. In this study, we established a HPLC-UV method for

**Table 1** Within–run variations and accuracy of the HPLC method for quantitation of phenylalanine (n = 3 )

Nominal Added Concentration (µg/ml)	Run Number	Measured Concentration (µg/ml)	Mean ± SD	CV%	Accuracy	Mean ± SD
0.5	1	0.45	0.53 ± 0.073	14	90.2	105 ± 14
	2	0.53			105.2	
	3	0.60			119.6	
2.5	1	2.83	25.62 ± 3.55	13.86	113.36	102.48 ± 14.18
	2	2.69			107.64	
	3	2.16			86.44	
10	1	11.34	10.29 ± 1.12	10.89	113.4	102.9 ± 11.2
	2	10.43			104.3	
	3	9.11			91.1	

**Table 2** Between–run variations and accuracy of the HPLC method for quantitation of phenylalanine (n = 3)

Nominal Added Concentration (µg/ml)	Run Number	Measured Concentration (µg/ml)	Mean ± SD	CV%	Accuracy	Mean ± SD
0.5	1	0.68	0.65 ± 1.01	15.61	136.02	129.40 ± 20.08
	2	0.73			145.02	
	3	0.53			106.80	
2.5	1	2.83	2.73 ± 0.25	9.34	113.36	109.16 ± 10.21
	2	2.44			97.52	
	3	2.92			116.60	
10	1	10.28	10.75 ± 0.44	4.12	102.8	107.5 ± 4.4
	2	11.16			111.6	
	3	10.81			108.1	

**Table 3** Relative recovery of phenylalanine by the HPLC method (n = 3)

Nominal Added Concentration (µg/ml)	Recovery (%)	Mean ± SD
0.5	82.65	84.97 ± 1.25
	80.92	
	75.00	
2.5	81.89	84.17 ± 1.61
	70.51	
	85.09	
10	82.78	96.68 ± 9.99
	75.34	
	87.20	

simultaneously detecting Phe levels in human plasma. The outcomes showed that our HPLC-UV method fully satisfied these conditions as mentioned above. For this method, the plasma preparation for analysis consist of a protein precipitation method. The validation tests on the developed method showed acceptable degree of linearity, sensitivity, precision, accuracy and recovery for the method.

### Conclusion

Briefly, we have successfully recognized a HPLC-UV method for simultaneously identifying Phe in human plasma. This method delivers a quick and a cheap alternative to those using HPLC with pre- or

post-column derivatization. A simple HPLC method was established and indorsed for Phe assay in plasma. The method was used successfully for quantization of Phe in plasma samples. System suitability tests showed that the established method is of suitable separation efficiency and peak shape.

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### Conflict of interest

Authors had declared no conflict of interest

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