

Review Article :

A Review on *Cyperus rotundus*: Ancient Weed to Modern Elixir of Life Phytochemistry and Therapeutic Uses of *Cyperus rotundus* (Mustaka)



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ABSTRACT

Background: Since time immemorial, humans have identified several herbs to treat various ailments. With the advancement of science and state-of-the-art technologies, different herbal extracts and chemical constituents of herbs were identified as therapeutic targets. *Cyperus rotundus*, also called mustaka, is one of the most ancient herbs widely distributed in tropical and subtropical regions across the globe. The tuberous and aerial parts of the herb were identified to possess various pharmacological properties.

Objectives: This review focuses on the various phytochemicals of mustaka and how these compounds exert pharmacological effects and their mode of action. The molecular and cellular effects of mustaka were also discussed based on the preclinical and clinical reports available using an array of in vitro, in vivo, and ex vivo methodologies.

Methods: The information from Google Scholar, Science direct, PUBMED, were reviewed with a special focus on the mode of action of *C. rotundus* from the data on animal and pre-clinical experiments to treat various diseases.

Conclusion: Based on the literature available on *C. rotundus* in Google Scholar, Science Direct, and PubMed, the pharmacological properties of mustaka were reviewed with a particular focus on its neuropharmacological activities. The mode of action of *C. rotundus* and its bioactive metabolites at the molecular biology level were demonstrated based on animal and preclinical experiments to cure various ailments. These diverse effects prove *C. rotundus* as a valuable traditional medicine for treating various disorders.

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Introduction

Herbal extracts and their chemical components are extensively studied to identify their various pharmacological effects. These studies advanced the effective utilization of herbal extracts as a traditional and alternative medicine to cure various ailments due to their fewer or no side effects. The commercialization and popularization of herbal extracts have led to their wide utilization as an ayurvedic medicine to treat various disorders. However, there is still a surge in the identification and commercialization of herbal medicines with no genotoxic and cytotoxic effects for treating various diseases. *Cyperus rotundus* is a weed distributed across tropical and subtropical regions, including India, Africa, Tunisia, and other countries. *C. rotundus* is one of the oldest medicinal herbs used from the age of Charaka to cure various diseases (Figure 1) [1]. In ancient times, *C. rotundus* powder was used as a flavoring agent on cooked meat, and the starch of the tuber was extracted to prepare noodles [2]. Recent studies also incorporated *C. rotundus* in minced beef meat to improve the quality, storability, and safety [3].

Several studies previously reported that *C. rotundus* possess anti-bacterial, anti-fungal, and mosquito-repellent activities [4, 5]. *C. rotundus* possesses various pharmacological properties, such as anti-inflammatory, anti-obesity, anti-diabetic, and neuroprotective effects [4, 6-8]. Our group also reported that *C. rotundus* possess strong free radical scavenging, anti-oxidant, and lipid peroxidation properties using an array of in vitro anti-oxidant and free radical scavenging activities. We also studied the in vivo anxiolytic and cognitive effects of *C. rotundus* in animal models of anxiety and hypobaric hypoxia in mice and rats, respectively [9, 10]. The oxide-nitrosative and anti-apoptotic effects of *C. rotundus* against hydrogen peroxide (H₂O₂), an oxidative stress insult and 3-morpholinosydnonimine (SIN-1), a nitrosative stress insult induced cytotoxicity were reported from our lab using SH-SY5Y cells, a widely used human cell line model to study neuronal stress/neuroprotective effects [11, 12]. Besides, we reviewed the ex-vivo anti-oxidant and antihemolytic effects of *C. rotundus* using H₂O₂ induced oxidative stress in White Blood Cell (WBC) and 2,2'-azobis(2-amidinopropane) dihydrochloride (AAPH)-induced hemolysis in WBC [9, 13].

However, little is known on the mode of action of *C. rotundus* and its chemical constituents on various ailments at the molecular level. Hence in the present review article, we discuss the diverse pharmacological effects of

C. rotundus and its phytochemicals in treating various diseases based on in vitro, in vivo, and ex vivo studies using animal, cell culture, and clinical studies. Here we discuss the interaction and mode of action of *C. rotundus* in regulating these ailments at the molecular level.

Cyperus rotundus taxonomical classification and cultivation

Cyperus rotundus L. (family: Cyperaceae), commonly known as musta, mustaka, nutgrass, java grass, purple nutsedge, red nutsedge [14], is native to India, Africa, southern and central Europe, and southern Asia [15]. It is widely distributed in tropical and subtropical regions worldwide [16]. *C. rotundus* is listed by Theophrastus [17] among other perfume plants of which “the most excellent and most fragrant come from Asia and sunny regions” (Historia Plantarum, IX 7.3). The genus name *Cyperus* is derived from the ancient Greek name Cypeiros, and the species name *rotundus* is from the Latin word round, which denotes the plant tuber [18]. It is a perennial sedge with umbel inflorescence, and fibrous roots reproduce widely through rhizomes and tubers. It is a widespread species under unfavorable conditions and is today considered the most troublesome weed in agriculture due to its competitive nature for ground nutrients with herbicide tolerance and high adaptability, which cause large yield losses [19]. However, it possesses various ethnomedicinal uses and therapeutic properties comprising essential oil and other phytochemical constituents. The most common targets of mustaka and its constituent include transcription factors, enzymes, growth factors, cytokines, kinases, proliferative factors, inflammatory mediators, receptors, and proteins involved in cell survival, apoptosis and metastasis (Figure 2).

Phytochemical Analysis of C. rotundus

Phytochemical analysis of herbal extracts has been investigated by various analytical techniques using sophisticated instruments, such as gas chromatography-mass spectrometry (GC-MS), high performance liquid chromatography (HPLC), and liquid chromatography-mass spectrometry (LC-MS) [20, 21]. The major phytochemicals of *C. rotundus* include essential oils, flavonoids, terpenoids, sesquiterpenes, sitosterol, cyperene, cyperol, nootkatone, and valencene [7, 22]. Sesquiterpene alkaloids, such as rotundines A, B, and C, were isolated from *C. rotundus* [23]. Chen et al. [24] analyzed the active constituents of *C. rotundus* by GC-MS, which shows the presence of α -cyperone. The phenolics and flavonoids are other groups of secondary metabolites which possess anti-oxidants and free radical scavenging activities

[25]. *C. rotundus* possesses phenolic compounds, such as 3-hydroxy-4-methoxy-benzoic acid, galloylquinic acid, ferulic acid, and flavonoid compounds, such as quercetin, luteolin, afzelechin, catechin [26]. Qualitative tandem liquid chromatography quadrupole time of flight mass spectrometry (LC-Q-TOF/MS) analysis of *C. rotundus* hydroethanolic fraction showed the presence of scaberin, auresidin, cyperol, 4,6,3,4-tetramethoxyaurone, luteolin 5-methyl ether, epoxycaryophyllane-5 α ,15-diol, 4-hydroxybenzoic acid, alpha cyperone, 4-7,-dimethyl-1-tetralone, p-coumaric acid, *Cyperusol* D, sulfuretin, juvenile hormone III, rotundine B, isokubosone, rotundine, beta rotunol, *Cyperusol* B2, *Cyperusol* A1, 6,3,4,-trihydroxy-4-methoxy-5-methylaurone, dolabella-3,7,18-triene, whereas Total Oligomeric Flavonoid fraction (TOF) fraction showed the presence of 4,6,3,4-tetramethoxyaurone, rotundine B, eudesma-2,4(15)-11-triene, *Cyperusol* D, γ -calacorene, 4,7-dimethyl-1-tetralone, β -rotunol, isocorymbolone [13, 27].

Phytochemicals of essential oils of *Cyperus rotundus* are as follows: monoterpene hydrocarbons; camphene, limonene, p-cymene, sabinene, α -pinene, β -pinene, oxygenated monoterpenes derivatives; 1,8-cineole, terpinen-4-ol, α -terpineol, carvacrol, borneol, myrtenol, terpinolene, thymol, oxygenated sesquiterpenes derivatives; α -cadinol, (2E, 6E)-farnesol, α -cyperone, T-muurolol, elemol, mustakone, caryophyllene oxide, cubenol, T-cadinol, eugenol, monoterpene aldehydes; myrtenal, monoterpene ketones; carvone, dihydrocarvone, sesquiterpene hydrocarbon; α -copaene, aromadendrene, β -elemene, valencene, (E,E) α -farnesene, α -humulene, β -caryophyllene, γ -elemene, sesquiterpene ketones; nootkatone, sesquiterpene aldehyde; cinnamaldehyde, flavonoids; luteolin 3'-methyl ether, luteolin 7,3'-dimethyl ether, luteolin 5,3'-dimethyl ether, luteolin 7-glucuronide, luteolin 4'-glucoside, orientin, quercetin, quercetin 3-rutinoside, kaempferol, triterpenoid; n-tritriacontan-16-one, n-pentadecanyl-9-octadecenoate, n-tetradecanyl-n-octadec-9, 12-dienoate, khellin, visnagin, ammiol, khellol- β -D-glucopyranoside, phenylpropanoids; isoaragoside, chionoside A, pungenin, salidroside, helioside C, alkaloids; rotundine A, rotundine B, rotundine C, phenolic acids; gallic acid, chlorogenic acid, caffeic acid, p-coumaric acid, loganic acid, ferulic acid, ellagic acid, iridoides, benzodihydrofurans, and other compounds; *rotunduside* A, *rotunduside* B, *rotunduside* C, *rotunduside*, 10-O-p-hydroxybenzoyltheviridoside, 10-O-vnilloyltheviridoside, 6''-O-(trans-p-coumaroyl)-procumbide, 6-O-p-coumaroylgenipin gentiobioside, 1-[2,3-dihydro-6-hydroxy 4,7-dimethoxy-2S-(prop-1-en-2yl)benzofuran-5-yl]ethanone, 1 α -methoxy-3 β -hydroxy-4 α -(3',4'-dihydroxyphenyl)-1,2,3,4 tetrahydronaphthalene,

1 α ,3 β -dihydroxy-4 α -(3',4'-dihydroxyphenyl) -1,2,3,4-tetrahydronaphthalene, n-butyl- β -D-fructopyranoside

ethyl- α -D-glucopyranoside, trans-(2-chlorovinyl) dimethylethoxysilane, 5-hydroxymethyl furfural, vanillin lactoside, 2-propenoic acid, 3-(4-hydroxy-3-methoxy phenyl)-methyl ester, 9, 12, 15-octadeca trienoic acid, 2, 3-bis [(trimethyl)oxy] propyl ester, methyl 3,4-dihydroxy benzoate, ipolamiide, 6b-hydroxyipolamiide, and rutin. The major phytochemicals of *C. rotundus* are presented in Table 1 with their classifications, molecular formula, and structures.

Neuro-pharmacological Effects of *C. rotundus*

The nervous system is highly vulnerable to the stress induced by external and internal stimuli that leads to neurodegeneration and cognitive decline. Various herbal extracts were determined as safe due to fewer side effects. They were evaluated by various preclinical methods and used across the globe from time immemorial because of their valuable properties [12, 28]. Understanding the mode of actions of different known exogenous and endogenous chemical agents or environmental agents that damage the nervous system or the herbal compounds or extracts that inhibit the neuronal damage is an interesting area of research. This line of research has strong potential in identifying novel neuro-pharmacological drugs to treat an array of neurodegenerative ailments. Various studies demonstrated the neuropharmacological effects of *C. rotundus* as presented in Table 2.

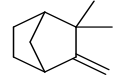
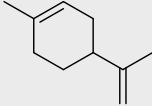
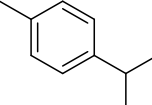
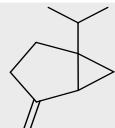
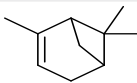
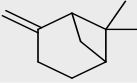
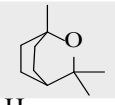
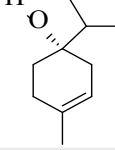
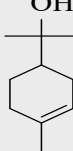
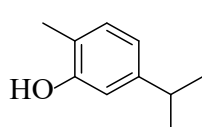
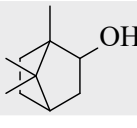
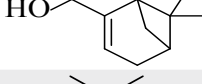
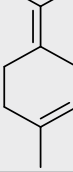
Anti-alzheimer activity of *C. rotundus*

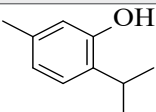
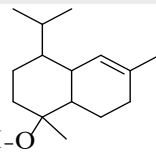
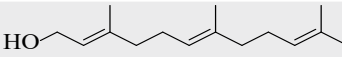
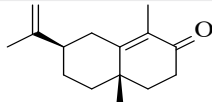
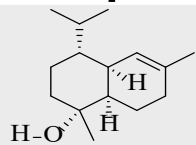
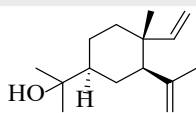
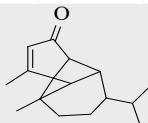
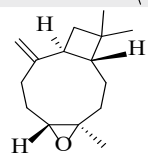
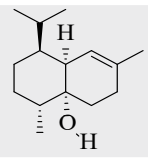
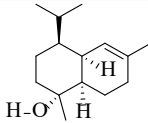
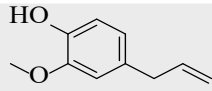
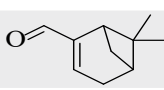
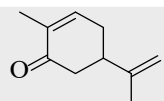
Alzheimer Disease (AD) is an irreversible, progressive disorder that leads to cognitive impairment and neurodegeneration. Amyloid plaques, microtubule fibers, and neurofibrillary tangles are observed in the brains of patients with AD. Oxidative stress is known to play a key role in the pathogenesis of AD [29]. The hydroalcoholic fraction of *C. rotundus* improved learning impairment, following the amyloid β peptide ($A\beta$) treatment-induced memory impairment in rats [30]. α -Cyperone, one of the major compounds of *C. rotundus*, binds and interacts with tubulin and destabilizes microtubule polymerization, and further reduces inflammation associated with AD [31].

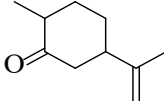
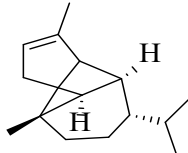
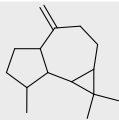
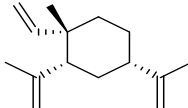
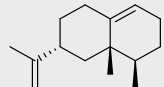
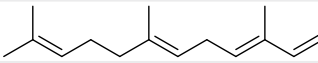
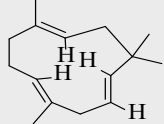
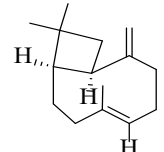
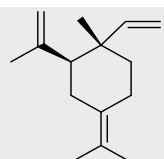
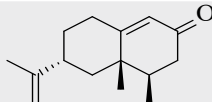
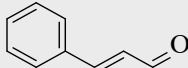
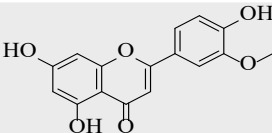
Anti-Parkinson Effects of *C. rotundus*

Parkinson disease is a progressive neurological disease in which degeneration of the dopaminergic neurons in the brain plays a key role in neurodegeneration. Lee et al. [32] reported the neuroprotective activity of water extract

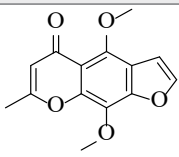
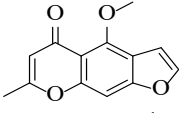
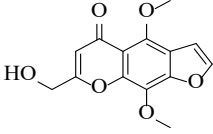
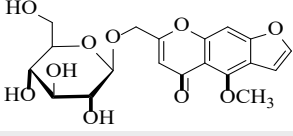
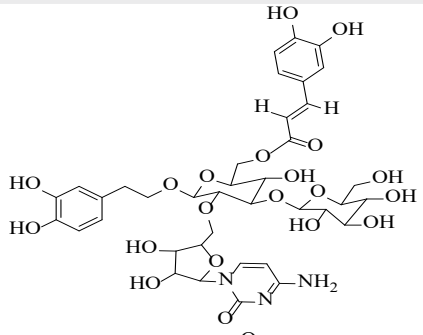
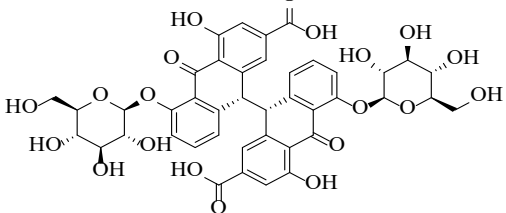
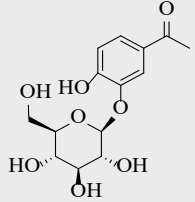
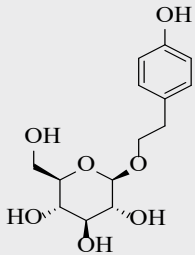
Table 1. Phytochemicals of cyperus rotundus according to their classification, molecular formula, and chemical structures

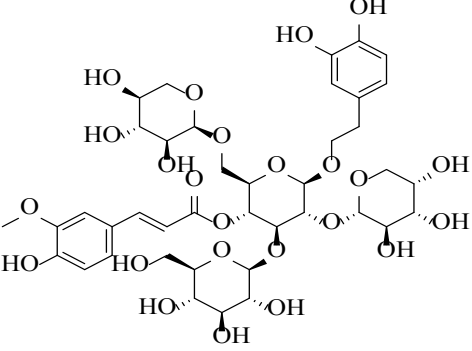
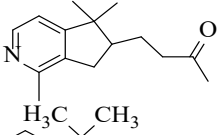
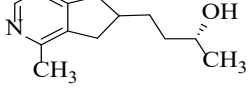
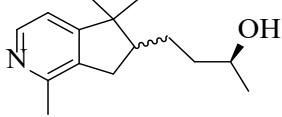
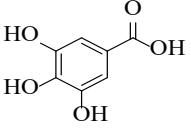
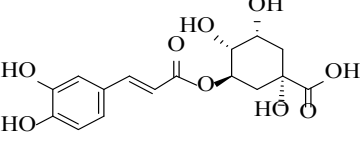
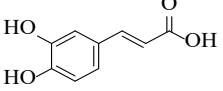
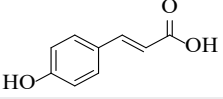
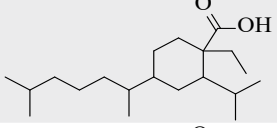
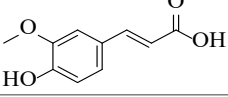
Phytochemicals of <i>C. rotundus</i>	Molecular Formula	Structure	References
Monoterpene hydrocarbons			
Camphene	C ₁₀ H ₁₆		[89]
Limonene	C ₁₀ H ₁₆		[90]
p-Cymene	C ₁₀ H ₁₄		[89]
Sabinene	C ₁₀ H ₁₆		[91, 92]
α-Pinene	C ₁₀ H ₁₆		[93, 94]
β-Pinene	C ₁₀ H ₁₆		[93]
Oxygenated monoterpenes derivatives			
1,8-Cineole	C ₁₀ H ₁₈ O		[95, 96]
Terpinen-4-ol	C ₁₀ H ₁₈ O		[97]
α-Terpineol	C ₁₀ H ₁₈ O		[98]
Carvacrol	C ₁₀ H ₁₄ O		[99]
Borneol	C ₁₀ H ₁₈ O		[100]
Myrtenol	C ₁₀ H ₁₆ O		[101]
Terpinolene	C ₁₀ H ₁₆		[102]

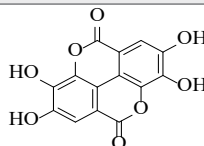
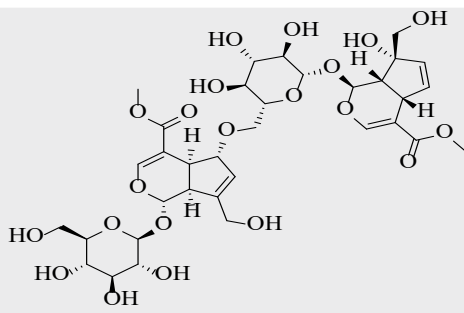
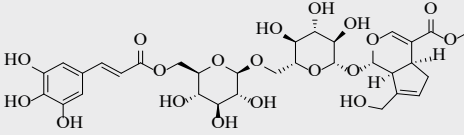
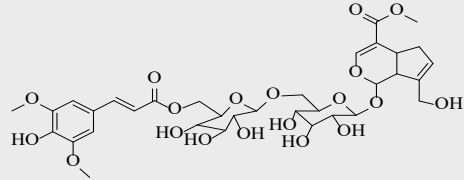
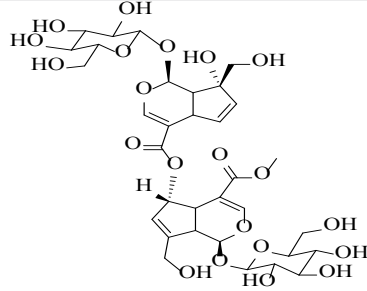
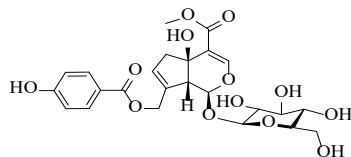
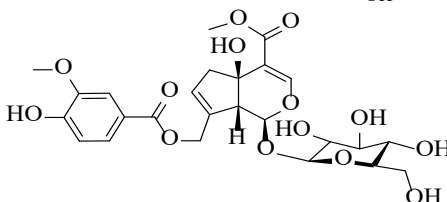
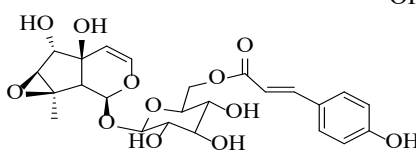
Phytochemicals of <i>C. rotundus</i>	Molecular Formula	Structure	References
Monoterpene hydrocarbons			
Thymol	C ₁₀ H ₁₄ O		[103, 104]
Oxygenated sesquiterpenes derivatives			
α-Cadinol	C ₁₅ H ₂₆ O		[105]
(2E,6E)-Farnesol	C ₁₅ H ₂₆ O		[106]
α-Cyperone	C ₁₅ H ₂₂ O		[107]
T-Muurolol	C ₁₅ H ₂₆ O		[108]
Elemol	C ₁₅ H ₂₆ O		[109]
Mustakone	C ₁₆ H ₂₂ O		[110]
Caryophyllene oxide	C ₁₅ H ₂₄ O		[111]
Cubenol	C ₁₅ H ₂₆ O		[112]
T-Cadinol	C ₁₅ H ₂₆ O		[113]
Eugenol	C ₁₀ H ₁₂ O ₂		[114]
Monoterpene aldehydes			
Myrtenal	C ₁₀ H ₁₄ O		[115]
Monoterpene ketones			
Carvone	C ₁₀ H ₁₄ O		[116, 117]

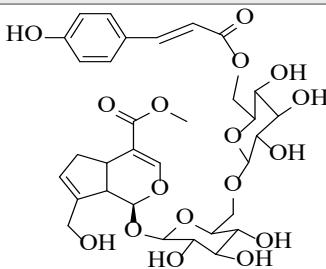
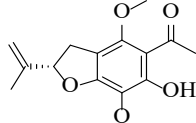
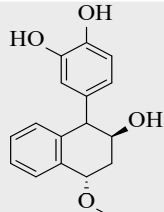
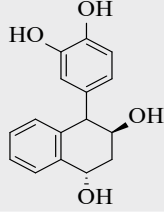
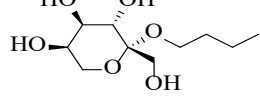
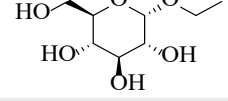
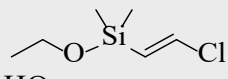
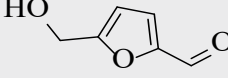
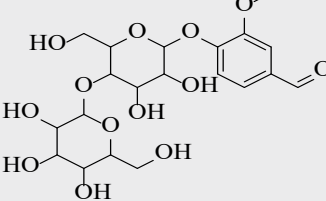
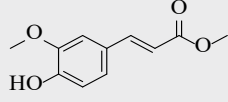
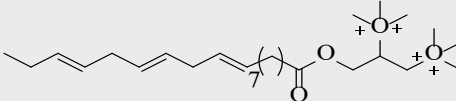
Phytochemicals of <i>C. rotundus</i>	Molecular Formula	Structure	References
Monoterpene hydrocarbons			
Dihydrocarvone	C ₁₀ H ₁₆ O		[118]
Sesquiterpene hydrocarbon			
α -Copaene	C ₁₆ H ₂₄		[119]
Aromadendrene	C ₁₅ H ₂₄		[120]
β -Elemene	C ₁₅ H ₂₄		[121]
Valencene	C ₁₅ H ₂₄		[7]
(E,E)- α -Farnesene	C ₁₅ H ₂₄		[122]
α -Humulene	C ₁₅ H ₂₄		[123, 124]
β -Caryophyllene	C ₁₅ H ₂₄		[125, 126]
γ -Elemene	C ₁₅ H ₂₄		[127]
Sesquiterpene ketones			
Nootkatone	C ₁₅ H ₂₂ O		[7, 128, 129]
Sesquiterpene aldehyde			
Cinnamaldehyde	C ₉ H ₈ O		[130, 131]
Flavonoids			
Luteolin 3'-methyl ether	C ₁₆ H ₁₂ O ₆		[132]

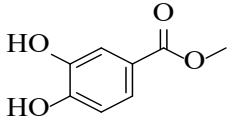
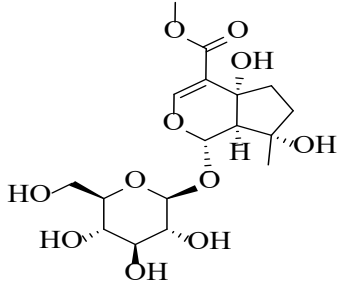
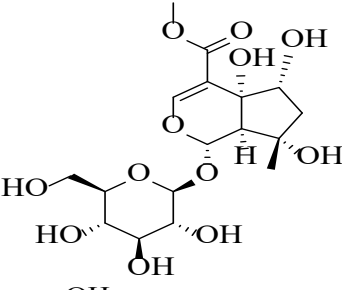
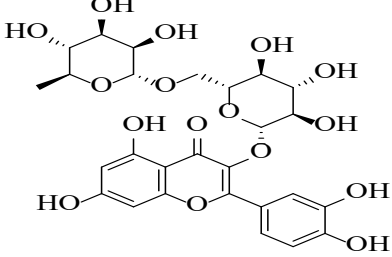
Phytochemicals of <i>C. rotundus</i>	Molecular Formula	Structure	References
Monoterpene hydrocarbons			
Luteolin 7, 3'-dimethyl ether	C17H14O6		
Luteolin 5, 3'-dimethyl ether	C17H14O6		[133]
Luteolin 7-glucuronide	C21H18O12		
Luteolin 4'-glucoside	C21H20O11		
Orientin	C21H20O11		[134]
Quercetin	C15H10O7		
Quercetin 3-rutinoside	C27H30O16		[132]
Kaempferol	C15H10O6		
Triterpenoid			
n-Tritriacontan-16-one	C33H66O		
n-Pentadecanyl-9-octadec-enoate	C33H63O2		[135]
n-Tetradecanyl-n-octadec-9, 12-dienoate	C32H59O2		

Phytochemicals of <i>C. rotundus</i>	Molecular Formula	Structure	References
Monoterpene hydrocarbons			
Khellin	C14H12O5		
Visnagin	C13H10O4		[133]
Ammiol	C14H12O6		
Khellol-β-D-glucopyranoside	C19H20O10		
Phenylpropanoids			
Isoaragoside	C38H47N3O20		[136]
Chionoside A	C42H38O20		
Pungenin	C14H18O8		[137]
Salidroside	C14H20O7		

Phytochemicals of <i>C. rotundus</i>	Molecular Formula	Structure	References
Monoterpene hydrocarbons			
Helioside C	C ₄₀ H ₅₄ O ₂₄		[136]
Alkaloids			
Rotundine A	C ₁₅ H ₂₁ N ₁ O		[23]
Rotundine B	C ₁₅ H ₂₃ N ₁ O		
Rotundine C	C ₁₅ H ₂₃ N ₁ O		
Phenolic acids			
Gallic acid	C ₇ H ₆ O ₅		[138]
Chlorogenic acid	C ₁₆ H ₁₈ O ₉		
Caffeic acid	C ₉ H ₈ O ₄		
p-Coumaric acid	C ₉ H ₈ O ₃		
Loganic acid	C ₂₀ H ₃₈ O ₂		
Ferulic acid	C ₁₀ H ₁₀ O ₄		[138, 139]

Phytochemicals of <i>C. rotundus</i>	Molecular Formula	Structure	References
Monoterpene hydrocarbons			
Ellagic acid	C ₁₄ H ₆ O ₈		[140]
Iridoides, benzodihydrofurans, and miscellaneous			
Rotunduside A	C ₃₄ H ₄₆ O ₂₁		[136]
Rotunduside B	C ₃₂ H ₄₀ O ₁₉		
Rotunduside C	C ₃₄ H ₄₄ O ₁₉		[137]
Rotunduside	C ₃₃ H ₄₄ O ₂₁		
10-O-p-Hydroxybenzoyltheviridoside	C ₂₄ H ₂₈ O ₁₃		[141]
10-O-Vnilloyltheviridoside	C ₂₅ H ₃₀ O ₁₄		
6''-O-(trans-p-Coumaroyl)-procumbide	C ₂₄ H ₂₈ O ₁₂		

Phytochemicals of <i>C. rotundus</i>	Molecular Formula	Structure	References
Monoterpene hydrocarbons			
6-O-p-Coumaroylgenipin gentiobioside	C32H40O17		[136]
1-[2,3-Dihydro-6-hydroxy-4,7-dimethoxy-2S-(prop-1-en-2yl) benzofuran-5-yl]ethanone	C15H18O5		[142]
1α-Methoxy-3β-hydroxy-4α-(3',4'-dihydroxyphenyl)-1,2,3,4-tetrahydronaphthalene	C17H18O4		[143]
1α,3β-Dihydroxy-4α-(3',4'-dihydroxyphenyl)-1,2,3,4-tetrahydronaphthalene	C16H16O4		
n-Butyl-β-D-fructopyranoside	C10H20O6		[140]
Ethyl-α-D-glucopyranoside	C8H16O6		
trans-(2-Chlorovinyl) dimethylethoxysilane	C6H13ClOSi		
5-Hydroxymethyl furfural	C6H6O3		
Vanillin lactoside	C20H28O13		[144]
2-Propenoic acid, 3-(4-hydroxy-3-methoxy phenyl)-methyl ester	C11H12O4		
9, 12, 15-Octadeca trienoic acid, 2, 3-bis [(trimethyl)oxy] propyl ester	C27H52O44+		

Phytochemicals of <i>C. rotundus</i>	Molecular Formula	Structure	References
Monoterpene hydrocarbons			
Methyl 3,4-dihydroxy benzoate	C ₈ H ₈ O ₄		
Ipolamiide	C ₁₇ H ₂₆ O ₁₁		
6b-Hydroxyipolamiide	C ₁₇ H ₂₆ O ₁₂		[145]
Rutin	C ₂₇ H ₃₀ O ₁₆		

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of *Cyperus* in an in vitro model of PC12 cells. The authors reported that 50 and 100 µg/mL doses of the extract restored 50% of cell death challenged by 6-hydroxydopamine, which exerts Parkinson-like effects by mediating oxidative damage. The extract efficiently reduced Reactive Oxygen Species (ROS), Nitric Oxide (NO) formation, and dissipated Mitochondrial Membrane Potential (MMP). Further, the extract downregulated caspase-3 expression, a biomarker of apoptosis, and effectively rescued the dopaminergic neuronal damage analyzed by tyrosine hydroxylase immunostaining, which is a biomarker of neurodegeneration suggesting the potential neuroprotective mechanism of the *Cyperus* extract.

Anti-anxiety activity of *C. rotundus*

Anxiety is a condition that every human experiences during their lifetime, but if it persists for a long duration, it develops as a mental illness that affects the psychological

well-being of an individual [33, 34]. Several drugs, including benzodiazepines that non-selectively target GABA_A receptors, are used as anxiolytics to treat anxiety [35]. However, several herbal remedies are also demonstrated as an anxiolytic [9, 34]. Our group demonstrated that hydroalcoholic extract of *C. rotundus* exhibits anxiolytic effects in a mouse model evaluated by an array of behavioral tests [27]. In a previous study, Sunil et al. [8] also demonstrated that the TOF fraction of *C. rotundus* reversed the anxiogenic behavior in rats and decreased the neurological deficits against cerebral ischemia-reperfusion injury.

Anti-depression property of *C. rotundus*

Depression is a psychological disease characterized by a sense of persistent sadness and or loss of interest. Natural mood enhancements and herbal supplements ease mild depression and are recommended as therapy. Wei et al. [36] evaluated the anti-depressive activity of

Table 2. Therapeutic, medicinal, and industrial applications of cyperus rotundus

Biological activity/ Application	Experimental Design	Results	References
In vitro anti-oxidant	Free radical scavenging assays like 2,2-diphenyl-1-picrylhydrazyl (DPPH), ABTS, superoxide, hydroxyl, nitric oxide, xanthine oxidase, hydrogen peroxide, and metal chelating, reducing power, lipid peroxidation activities	The ethanolic, methanolic, ethyl acetate, water, total oligomeric flavonoid, and oil fractions and active compounds such as stilbene dimers, cassigarol E, scirpusin A and B, cyperusphenol B, nootkatone, aristolone, solavetivone, ethyl-alpha-D-glucopyranoside (2), adenosine, (-)-(E)-caffeoylmalic acid, isovitexin, luteolin 7-O-β-D-glucuronopyranoside-6''-methyl ester, luteolin 4'-O-β-D-glucuronopyranoside, luteolin 7-O-β-D-glucuronopyranoside, uridine and of <i>C. rotundus</i> leaves, flowers and rhizomes exhibited potent free radical scavenging activities.	[9, 26, 59, 62-64, 86, 146-150]
Anti-bacterial	<i>Bacillus pumilis</i> , <i>S. pyogenes</i> , <i>Escherichia coli</i> , <i>Shigella flexneri</i> , <i>Staphylococcus aureus</i> , <i>Klebsiella pneumoniae</i> , <i>Enterobacter aerogenes</i> , <i>Salmonella paratyphi</i> , <i>Pseudomonas aeruginosa</i> , <i>Proteus vulgaris</i> , <i>Salmonella typhimurium</i> , <i>Salmonella typhimurium</i> , <i>Salmonella enteritidis</i> , <i>Enterococcus faecalis</i>	Rhizome / Hexane, Ethanol, Acetone, Aqueous, Methanol, Fruit/oil, Aerial parts, Lyophilized infusion, Total oligomeric flavonoids, Ethyl acetate	[57-61]
Anti-fungal	<i>Candida albicans</i>	Ethanol extract	[151]
Anti-viral	Herpes Simplex-1 Virus (HSV)	Tuber	[152]
Anti-malarial	African malaria vectors (<i>Anopheles gambiae</i> and <i>Anopheles arabiensis</i>)	Cedrol, a sesquiterpene alcohol	[153]
Anti-plasmodial	<i>Plasmodium falciparum</i>	Ethyl acetate	[154]
Ovicidal and larvicidal	<i>Aedes albopictus</i>	Tubers-essential oil	[155]
Mosquito repellent	<i>Anopheles culicifacies</i>	Hexane extract of the tuber	[4]
Insecticidal	<i>Blattella germanica</i>	Steam distillate constituents of <i>C. rotundus</i> (L.) rhizome	[156]
Anti-obese	The anti-obese property was verified using Obese Zucker rats and 3T3-F442 adipocytes	<i>C. rotundus</i> hexane fraction for 60 days exhibited a significant reduction in weight gain and stimulated lipolysis and thermogenesis by binding to β-adrenoreceptors in 3T3-F442 adipocytes	[6]
Hypolipidemic	Mechanism of amentoflavone on inhibition of uterine tumors.	Biflavone constituents of <i>C. rotundus</i>	[72]
α-Amylase and α-glycosidase inhibitory property	α-Glucosidase and α-amylase inhibitory activities were evaluated by in vitro enzyme inhibition assays which contribute to the antihyperglycemic activity.	Acarbose, tricrin, isorhamnetin, oreintin, vitexin, myricetin 3-O-β-D-galactopyranoside, isovitexin, luteolin 7-O-β-D-glucuronopyranoside, luteolin 4'-O-β-D-glucuronopyranoside, myricetin 3-O-β-D-galactopyranoside, protocatechuic acid, ellagic acid, chlorogenic acid, orientin, epiorientin, isorhamnetin, luteolin 7-O-β-D-glucuronopyranoside, luteolin 4'-O-β-D-glucuronopyranoside, (-)-(E)-caffeoylmalic acid, caffeic acid, p-coumaric acid, tricrin, salicylic acid vitexin, isovitexin, methanolic extract/ 3,4',5,6,7,8-hexahydroxyflavane, cassigarol E, scirpusin A and B	[140, 150]
Protein glycation, protein carbonylation, protein oxidation, and protein nitration inhibition	Fructose, AAPH, 3-morpholinodnonimine (SIN-1) mediated damage of bovine serum albumin (BSA) was verified.	Ethanolic extract of <i>C. rotundus</i> aerial part and rhizome inhibits fructose-induced protein glycation, protein carbonyl content, AAPH/SIN-1 mediated protein oxidation, protein nitration, and 3-nitrotyrosine.	[54, 86]
Anti-diabetic	The anti-diabetic activity was checked in alloxan-induced hyperglycemic rats	<i>C. rotundus</i> hydroalcoholic extract at 500 mg/kg oral administration for seven days reduced blood glucose levels.	[146]
Wound healing	Wound healing activity	Ethanol fraction of rhizome	[157]
Anti-mutagenic	<i>Salmonella typhimurium</i> assay, Antimutagenic activity against Aflatoxin and sodium azide in TA100 and TA1535 assay system	Total Oligomeric Flavonoid (TOF) and ethyl acetate and methanol extracts from aerial parts of <i>C. rotundus</i>	[62]
Anti-diarrheal	The anti-diarrheal activity was checked against castor oil-induced diarrhea in mice.	Methanol and petroleum ether fractions of <i>C. rotundus</i> exhibited anti-diarrheal activity	[80]
Anti-platelet	<i>C. rotundus</i> 70% extract and its active compounds were checked for anti-platelet activity by rat platelet aggregations in vitro and ex vivo, and on mice, tail bleeding assays.	Among the active compounds, nootkatone exhibited the most potent anti-platelet effects, followed by 4-Cymene, β-pinene, 1,8-cineole, limonene, valencene, caryophyllene oxide, and coumarin.	[128]

Biological activity/ Application	Experimental Design	Results	References
Anti-allergic	Sesquiterpenes and monoterpenes of <i>C. rotundus</i> were evaluated for anti-allergic activity in rat basophilic leukemia (RBL)-1 cells.	Sesquiterpenes inhibited 5-lipoxygenase production in RBL-1 cells and hexosaminidase release in antigen-stimulated RBL-2H3 cells. Valencene and nootkatone also inhibited the delayed-type hypersensitivity in mice.	[76]
Anti-inflammatory	MTT assay, real-time PCR assay, ELISA, western blot analysis, siRNA transfections of heme oxygenase-1(HO-1) and Nrf2 with murine microglial cell line BV-2, neuroblastoma cell line SH-SY5Y cells, and hippocampal cell line HT22 cells, InnoZyme TACE activity assay, in vitro anti-inflammatory activity using THP-1 monocytic cells and in vivo anti-inflammatory activity were confirmed by carrageenan-induced rat paw edema as a model.	α -Cyperone, ingredients of <i>C. rotundus</i> oil decreased the production of tumor necrosis factor- α (TNF- α), interleukin-6 (IL-6) and interleukin-1 β (IL-1 β) in LPS-induced BV-2 cells, anti-inflammatory activity in activated macrophage, α -cyperone inhibit PMA-induced endothelial cell protein C receptor (EPCR) shedding through PKC pathway, lupeol showed potent IL-1 β activity inhibition in THP-1 monocytic cells and also displayed significant in vivo anti-inflammatory activity, isocyperol, a sesquiterpene from rhizomes suppress septic shock through negative regulation of pro-inflammatory factors.	[66, 158-160]
Analgesic	Treatment of antipyretic and analgesic activities, acetic acid-induced mice stretching model, acetic acid-induced writhing assay	The crude extract of <i>C. rotundus</i> exhibited the dose-dependent spasmolytic effect in rabbit jejunum by inhibiting the spontaneous and K ⁺ (80 mM)-induced contractions, essential oil from fresh rhizome shows good analgesic effect. TOF-enriched extract	[24, 134, 161, 162]
Anti-proliferative	Thiazolyl blue tetrazolium bromide (MTT) assay	Terpene derivatives had anti-proliferative activity against the HCT116 cell line.	[163]
Anti-genotoxic	Chromosome aberrations assay	Aqueous, ethyl acetate, methanol, and TOF-enriched extracts exhibit no genotoxic activity	[161]
Anti-carcinogenic	(MTT) assay, migration assay in different cancer cell lines, electrophysiological recordings, western blot analysis, terminal deoxynucleotidyl transferase dUTP nick end labeling assay, caspases activity assay	Methanolic extract of <i>C. rotundus</i> rhizome (MRCr) showed significant anticancer activity, the effect of amentoflavone on inhibition of uterine tumors, valencene (1) from the rhizomes inhibits skin photoaging-related ion channels and UV-induced melanogenesis in B16F10 melanoma cells, 6-acetoxy cyperene (ethanol extract) from rhizome induce apoptosis in ovarian and endometrial cancer cells	[72-74]
Acetylcholinesterase (AChE) inhibition property	2,2-diphenyl-1-picrylhydrazyl (DPPH) and ferric reducing anti-oxidant power (FRAP) assays.	Ethanol extract of <i>C. rotundus</i> inhibits AChE.	[49]
Anti-convulsant	Tonic seizures induced by maximal electroshock (MES) and pentylenetetrazole (PTZ).	Ethanol extract significantly reduced hindlimb extension and convulsion duration in albino rats.	[14]
Neuroprotective	Morris water maze test (determination of spatial memory), western blot analysis, Nissl staining	Enhanced memory increased neuronal density, with activated pERK1/2 were observed in the hippocampus of an extract of <i>C. rotundus</i> and <i>Zingiber officinale</i> (CP1) treated rats. Cyperus ethanolic extract (CRE) constituents of scirpusin A and scirpusin B have therapeutic use against neurodegenerative diseases. EECR prevents pyramidal cell loss in the CA1 region of the hippocampus.	[11, 48, 49, 54, 164]
Sedative, anti-anxiety, anti-depressant	Sesquiterpenes of <i>C. rotundus</i> were examined by radiobinding assay using rat cerebrocortical brain slices. Anti-depressant effect of a polyherbal formulation Yueju-Wan extract composed of <i>C. rotundus</i> was checked for its anti-depressant activity in mice model by locomotor activity, tail suspension, and forced swim tests. The anxiolytic activity of 70% ethanol fraction of <i>C. rotundus</i> was checked by behavioral tests, such as open field, light-dark box, elevated plus maze, and Vogel conflict tests.	Ethyl extract fraction isolated sesquiterpene, isocurcumenol was found to be benzodiazepine receptor agonist explaining the sedative effects of <i>C. rotundus</i> . Supplementation of mice with Yueju-Wan extract showed anti-depressant effects. Oral administration of mice with <i>C. rotundus</i> at 200 mg/kg showed anxiolytic effects.	[9, 36, 46]
Flooding tolerance	<i>C. rotundus</i> grown in upland and low land ecosystems were compared in terms of growth habits, carbohydrate reserves and metabolism, and activities of enzymes involved in alcoholic fermentation, such as alcohol dehydrogenase and pyruvate decarboxylase.	The study demonstrates that the enzyme activities and soluble sugars were regulated, favoring the high growth of the weed <i>C. rotundus</i> in lowland ecosystems than uplands in a hypoxic environment which poses a major threat to rice fields.	[165]

Biological activity/ Application	Experimental Design	Results	References
Phytoremediation property	Bioaccumulation coefficient as a measure of phytoremediation, total petroleum hydrocarbon (TPH)	Petroleum hydrocarbons contaminated sites, phytoremediation of chromium by aquatic and terrestrial weeds	[166-168]
Food application	Supercritical fluid extraction with carbon dioxide	The optimum yield of <i>C. rotundus</i> oil (1.82%) was found under the following conditions: temperature, 37.6°C; pressure, 294.4 bar; extraction time, 119.8 min; and CO ₂ flow rate 20.9 L/h.	[169]

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the Chinese poly-herbal formulation Yueju-Wan (which possesses *Cyperus rotundus*) by behavioral analysis in a mice model. The extract efficiently reduced the immobility time analyzed by tail suspension and forced swimming tests but did not significantly affect locomotor activity. These outcomes may result from the active components of the herbal formulation, among which α -cyperone is the one suggesting the possible anti-depressive activity of *Cyperus* extract.

The neurological properties of the ethanolic extract of *Cyperus* were evaluated by Pal et al. [37] in a mice model. The authors calculated the LD50 of ethanolic extract as 240 mg/kg, IP. The sleeping time was potentiated maximally with 80 mg/kg of the extract and hypnotics pentobarbitone, diazepam, and meprobamate and exhibited analgesic activity and further potentiated the analgesic activity induced by morphine and pethidine. Additionally, the extract efficiently increased the survival time, decreased the mortality rate against strychnine and leptazol-induced convulsions, and also reduced behav-

ioral reflexes suggesting the potential anti-depressive action of the *Cyperus* extract.

Anti-epileptic/anti-convulsant activity of *C. rotundus*

Epilepsy is a group of seizures characterized by abnormal recurrent and spontaneous electrical discharge of cerebral neurons, and its control requires continuous anti-convulsant medication. Khalili et al. [38] described the neuroprotective property of hydroalcoholic extract of *Cyperus* in mice model by pentylenetetrazole (PTZ-induced kindling), which mimics epilepsy-induced seizures in rats and mice. The hydroalcoholic extract of *C. rotundus* efficiently ameliorated the PTZ-induced seizure formation and reduced epilepsy-associated neuro-oxidative stress and restored the anti-oxidant status of the brain in an in vivo model of epilepsy [38, 39].

Sedative activity of *C. rotundus*

Sedatives or tranquilizers act on the nervous system and induce sleep by reducing irritability or excitement. The sedative and analgesic effect of *C. rotundus* with its essential oil and crude extract was evaluated at a dose of 300 and 500 mg/kg bodyweight by tail-flick method on mice [24, 40]. Dilipkumar et al. [41] showed that *C. rotundus* extract has an analgesic effect and significantly potentiated the sleeping time of mice induced by standard hypnotics (diazepam). The sedation action might be due to enhancing brain serotonin and GABAergic transmission [42].

Anti-nociceptive activity of *C. rotundus*

The body contains nociceptive receptors that recognize neuropathic or nociceptive pain usually caused by an injury, physical pressure, or inflammation of some part of the body. Hydro-methanolic extract of *C. rotundus* exhibits anti-nociceptive activity and inhibits thermal and chemical-induced pain in mice. A study also demonstrated that *C. rotundus* did not cause any mortality,



Figure 1. *Cyperus rotundus*

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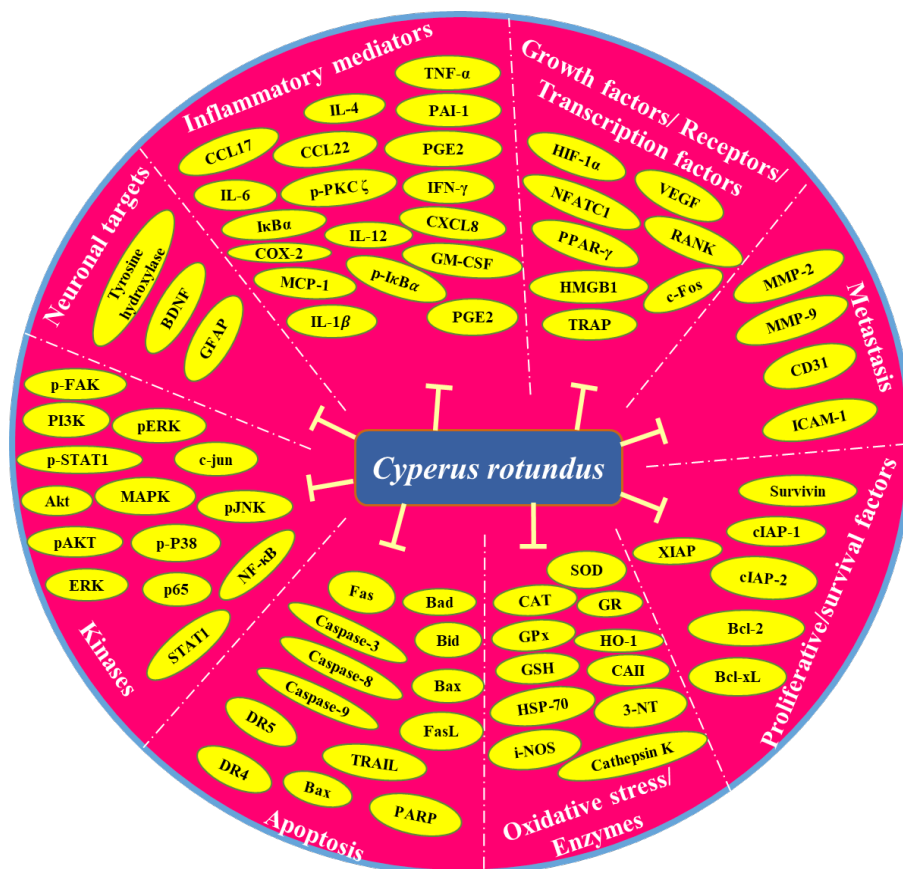


Figure 2. Molecular Targets of *Cyperus rotundus* and Its Bioactive Compounds

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behavioral changes, or allergic reactions and is safe up to 3000 mg/kg body weight in mice [43].

Nootropic effects of *C. rotundus*

The medications that exert neurological relief/neuro-protection and aid in the psychological well-being of individuals are considered nootropics [44, 45]. In an earlier study, Ha et al. [46] demonstrated the γ -aminobutyric acid (GABAA)-benzodiazepine (BZR) receptor activity of *Cyperus* extract. Activation of BZR, a key component of the GABA receptor complex, enhances the actions of GABA, an inhibitory neurotransmitter, on the Cl⁻ conductance of the neuronal membrane. The authors demonstrated the rat cerebrocortical BZR receptor agonistic activity of methanol, ethyl acetate fractions, and isourcemenol from ethyl acetate fraction by receptor binding assay.

Lee et al. [47] reported the neuroprotective activity of Ondamtanggamibang (ODG), a traditional Korean herbal remedy that encompasses 15% of *Cyperus* extract by using in vitro model of PC12 cells. About 600 μ g/mL of ODG restored cell viability with hydrogen peroxide and 6-hydroxydopamine (6-OHDA) challenge, respec-

tively, reducing ROS formation. The HO-1 gene expression is up-regulated with an increasing concentration of ODG. In contrast, zinc protoporphyrin IX (ZnPP-IX), an inhibitor of heme oxygenase-1(HO-1), the treatment showed poor cell viability against H₂O₂ challenge with ODG treatment. Also, cycloheximide and actinomycin-D inhibited ODG induced HO-1 expression, suggesting the possible protective role of ODG in oxidative stress via HO-1 expression.

In an elegant study, Sunil et al. [8] described the neuroprotective activity of Total Oligomeric Flavonoids (TOF) of *Cyperus* by in vivo model of cerebral-ischemia reperfusion injury (IR). In this study, 200 mg/kg TOF extract alleviated the neurological dysfunction and sensorimotor performance compared with IR rats. The extract also decreased the brain glutamate, glutamine synthetase activity and increased the Na⁺K⁺ATPase activity, thus reducing the excitotoxicity, a key event in neuronal dysfunction. The extract showed potential anti-anxiety activity evaluated by rats' behavioral functions and protected the degeneration of neurons as analyzed by histopathological sectioning of the brain. Furthermore, TOF treatment restored the anti-oxidant status of the IR brain,

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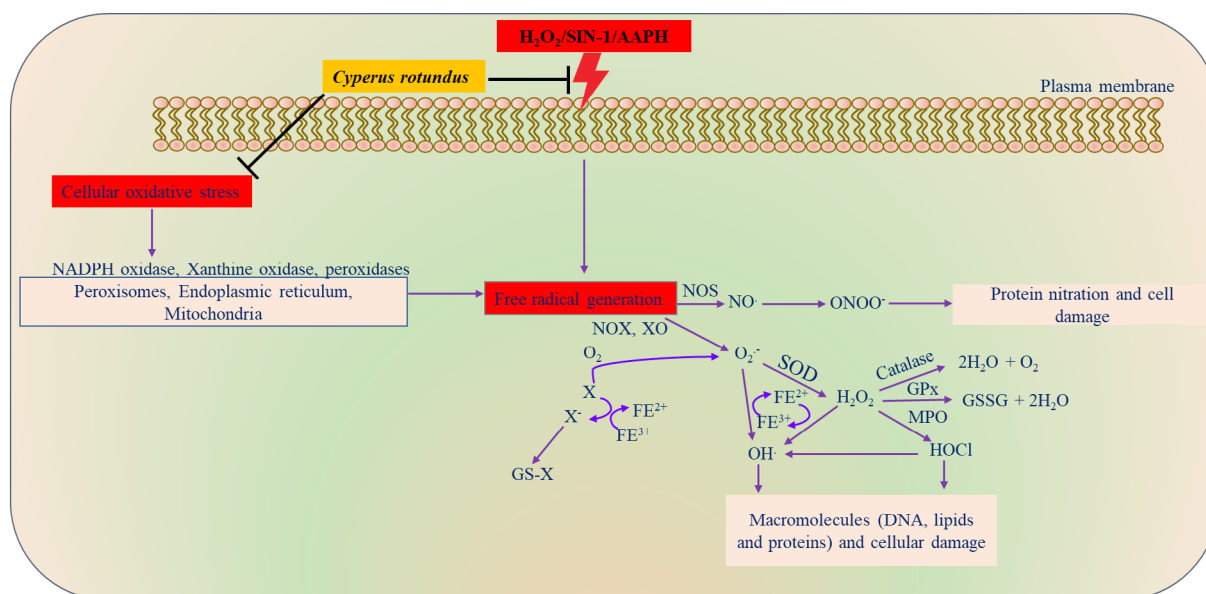


Figure 3. *Cyperus rotundus* Inhibiting Oxide-nitrosative Stress in Nervous System

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Extrinsic and intrinsic free radicals mediate oxidative-nitrosative stress, such as NADPH oxidase, xanthine oxidase, myeloperoxidase, peroxidases due to peroxisomes, endoplasmic reticulum. Also, mitochondria stress induces macromolecules, such as DNA, lipid, and protein damage and mitochondrial ROS to mediate neurodegeneration.

explaining the potential role of flavonoids in neuroprotection.

Memory impairment modulating activity of *C. rotundus*

Neurodegeneration as a consequence of neurological diseases such as Alzheimer, Parkinson, or due to ischemia, hypobaric hypoxia impairs the cognitive ability of individuals by damaging CA1 and CA3 regions of the hippocampus. These areas play a crucial role in memory and recognition ability. Various preclinical mice/rat models determined that *C. rotundus* administration inhibits neurodegeneration and memory impairment and prevents pyramidal cell loss in CA1 and CA3 regions of the hippocampus. Ethanolic extract of *C. rotundus* prevents ischemia, hypobaric hypoxia, and amyloid peptides-induced cognitive decline in rat and mice models, which was evaluated by Morris Water Maze Test [10, 30, 48-50]. Besides, the combined extract of *C. rotundus* and *Zingiber officinale* enhanced cholinergic function and decreased neurodegeneration and oxidative stress in a rat model of spatial memory impairment [49].

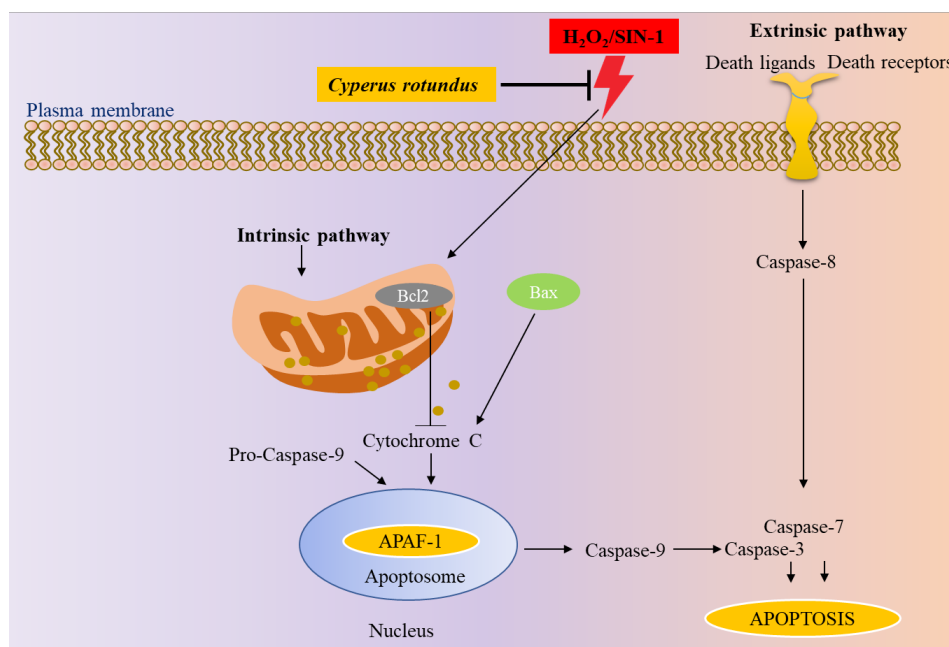
Acetylcholinesterase (AChE) inhibitory activity of *C. rotundus*

Acetylcholinesterase Inhibitors (AChEIs) inhibit the enzymatic elimination of acetylcholine, increasing its concentration at the postsynaptic membrane. AChEIs

are beneficial in improving cholinergic system deficits and are used as cognitive enhancers in AD. Sharma and Gupta [51] reported the AChE inhibitory activity of the methanolic extract of *Cyperus*. The extract at 0.5 mg/mL showed 50% inhibition of electrophorus electricus (electric eel) AChE. It also strongly inhibited AChE from leaves of wheat rather than tomato. The extract also inhibited seed germination, root and shoot length. These properties explain the potential neuroprotective activity and a war against plants and herbivore animals growing in the same habitat.

Neuro-oxide-nitrosative and neuronal apoptosis inhibitory properties of *C. rotundus*

Oxide-nitrosative stress and apoptosis are the central phenomena of various diseases, including nervous system-associated illnesses [52, 53]. The brain is a vital organ in the biological system and is highly susceptible to oxidative-nitrosative stress and apoptosis. Various reports demonstrated the anti-oxidant and anti-apoptotic effects of *C. rotundus* and its metabolites. We also observed that *C. rotundus* hydroalcoholic fractions exhibit neuro oxide-nitrosative and apoptosis inhibitory properties against H_2O_2 /SIN-1 induced oxidative-nitrosative stress and apoptosis in vitro in SH-SY5Y cells and restore anti-oxidant decline, inhibits ROS, inducible Nitric Oxide Synthase (iNOS), heat shock protein 70 (Hsp70), an anti-oxidant and anti-apoptosis protein expression, DNA damage and mitochondrial membrane potential [11, 54] (Figures 3 and 4). In another study, we demonstrated that the TOF fraction of *C. rotundus* also inhibited



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Figure 4. *Cyperus rotundus* Inhibiting H₂O₂/SIN-1-Mediated apoptosis and Mitochondrial Damage, Intrinsic and Extrinsic Apoptosis The step that commits cells to undergo apoptosis is permeabilization of the mitochondrial outer membrane (MOMP), decrease in mitochondrial membrane potential, and increase in mitochondrial ROS and neurodegeneration.

in vivo hypobaric hypoxia-induced neuronal ROS, LDH (lactate dehydrogenase) release, and improved anti-oxidant decline, and regulated HIF-1 α (Hypoxia-inducible factor 1-alpha), vascular endothelial growth factor (VEGF), and glial fibrillary acidic protein (GFAP) expression in vivo [55] (Figure 5).

Other Pharmacological Properties of *C. rotundus*

Anti-bacterial Property

Bacterial infections pose a risk to humans, and anti-bacterial agents suppress the growth and the ability of bacteria to reproduce [56]. Sini and Malathi [57] reported that hexane and water extracts of *Cyperus* tubers are potent inhibitors against *Bacillus pumilis* as determined by the zone of inhibition. Nima et al. [58] reported the antimicrobial activity of the oil extracted from fruits of *Cyperus* against Gram-positive bacteria, such as *Staphylococcus aureus*, compared to Gram-negative bacteria. The extract consists of cyperol, caryophyllene, cyperene, rotundine, and cyperone, which may be the active constituents responsible for the anti-bacterial activity of *Cyperus* oil extract. Kilani et al. [59] reported the anti-bacterial activity of different constituents of rhizome extract of *Cyperus*, which was further verified by Kilani-Jaziri et al. [60] in terms of both MIC (minimum inhibi-

tory concentration) and MBC (minimum bactericidal concentration) assays. Tambekar et al. [61] reported the anti-bacterial activity of *Cyperus* rhizome extract by the zone of inhibition assay.

Anti-oxidant Property

Several diseases are associated with oxidative stress, and anti-oxidant supplements/herbs inhibit/delay the disease progression and ameliorate the ROS-induced injury [13, 27]. Kilani et al. [62] reported that ethyl acetate, methanol, and TOF fractions of *Cyperus* tuber extract are potent scavengers of 2,2-diphenyl-1-picrylhydrazyl (DPPH) radicals. Nagulendran et al. [63] reported the anti-oxidant activity of ethanolic extract of *Cyperus* tubers by in vitro free radical scavenging assays. The extract efficiently scavenged superoxide, nitric oxide, hydroxyl radicals and also inhibited the lipid peroxidation induced by ferrous ions. These activities may account for the high polyphenolic content of the *Cyperus* extract. Kilani et al. [64] reported that the TOF fraction from tubers of *Cyperus* is a potent inhibitor of superoxide radicals than other fractions. In contrast, Jazari et al. [60] reported that ethyl acetate fraction is the most potent scavenger of superoxide radical among other fractions from aerial parts of *Cyperus*. These anti-oxidant activities were attributed to the high content of polyphenols, flavonoids, tannins, and sterols.

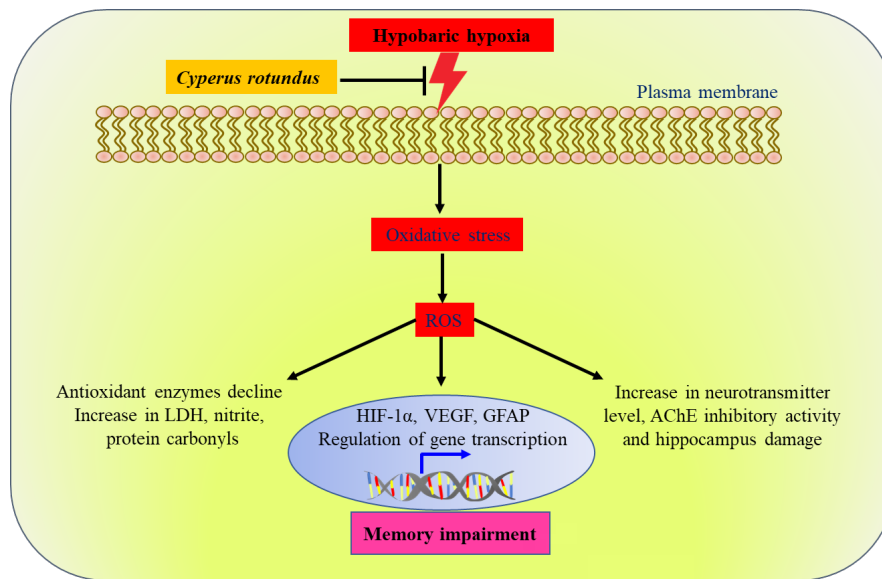


Figure 5. Hypobaric Hypoxia-Induced Oxide-Nitrosative Stress Mediating the Cognitive Impairment Modulating Effects of *C. rotundus*.

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Anti-platelet property

The clumping of platelets in the blood leads to blood clots and coagulation, which is commonly seen in injury and is critical for hemostatic plug formation and thrombosis. However, platelet aggregation is also seen in cardiovascular diseases. Seo et al. [65] described the anti-platelet activity of *Cyperus* ethanolic extract (CRE) and its active compound nootkatone in an in vitro model of rat platelet aggregation and ex vivo mice tail bleeding model. CRE extract at 300 µg/mL inhibited the platelet aggregation challenged by collagen, thrombin, and arachidonic acid. Nootkatone at 30 mg/mL inhibited isolated rat platelet aggregation and increased the bleeding time up to 3 folds suggesting the potential application of the extract in arterial thrombosis.

Anti-diabetic property

Diabetes is one of the major leading causes of death worldwide. It is due to impairment in insulin production by the pancreas that leads to abnormal metabolism of carbohydrates with elevation in glucose levels. Several herbal treatments are recommended for this chronic disease. Karkala et al. [66] reported the significant anti-diabetic activities by gallic acid and quercetin compounds from *C. rotundus*. In silico docking studies by Lydia and Sudarsanam [67] found a novel compound, 15-hydroxy-4-oxo-10-pentadecanoic acid lactone, that possesses anti-diabetic property. Singh et al. [68] evaluated the activity of ethanolic extract of *Cyperus rotundus* rhizomes

and showed a decrease in enzyme activities, cholesterol, and triglyceride levels in Streptozotocin (STZ)-induced diabetic mice.

Anti-obese and hypolipidemic property

Obesity is due to excessive fat and overweight, and such a condition puts the individual at higher risk for other diseases, such as type 2 diabetes, cardiovascular disease, and cancer. Bambhole et al. [69] reported the effect of *Cyperus* extract (CR) by in vitro and in vivo studies on adipose tissue metabolism. CR extract treatment alone significantly released Lipolytic Glycerol (LG) and Free Fatty Acids (FFA), but CR extract combined with epinephrine resulted in a two-fold increase of LG and FFA release from adipose tissue cells. Further oral administration of the extract to rats also released plasma glycerol and FFA and inhibited the release of plasma phosphate, an indicator of Adenosine Monophosphate Kinase (AMPk) mediated lipolytic pathway.

In an elegant study, Lemaure et al. [6] reported the anti-obese activity of hexane extract of *Cyperus* in obese Zucker rats and β-adrenergic receptor-expressing 3T3-F422 adipocytes, where the β-3AR activation leads to weight loss by decreasing fat level. The rats showed 2.7% and 4.2% decline in body weight without effect on food intake of the animals with 45 mg/kg and 220 mg/kg of hexane extract administration, respectively, for 60 days, and the extracts had no toxicity up to 1 g/kg body weight. The triglyceride levels were markedly increased

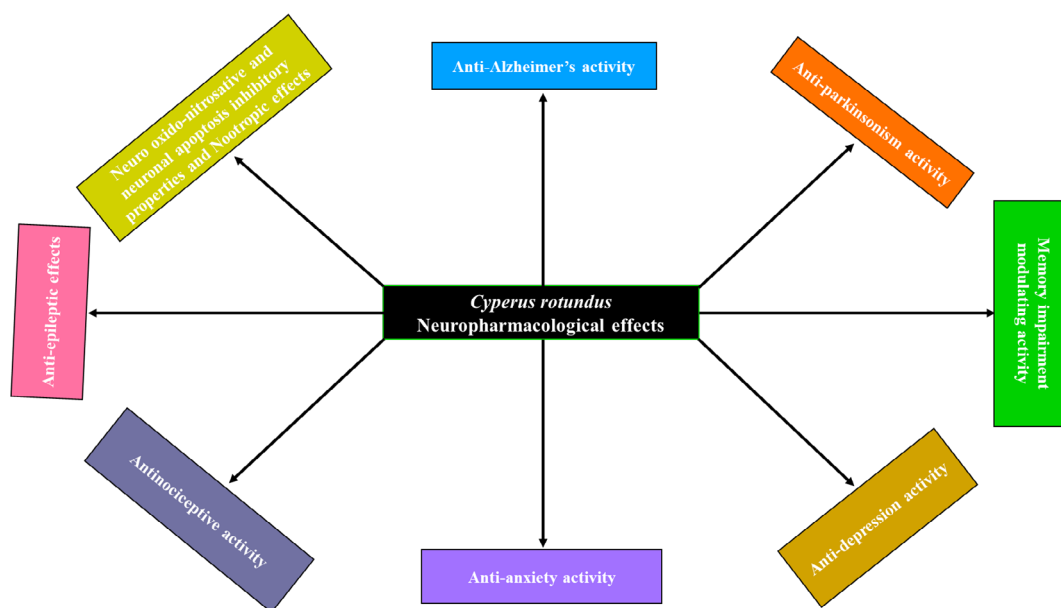


Figure 6. Graphical abstract: Neuropharmacological effects of *Cyperus rotundus*

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with a 220 mg/kg dose of extract. Further, the lipolytic activity of the extract was evaluated in adipocytes by measuring the β -AR binding assay by radiolabelling, whereas 250 $\mu\text{g/mL}$ of hexane extract showed a 2.5 fold increase in lipolytic activity as well as β -3AR activation leading to energy expenditure by brown adipose tissue thermogenesis showing the potential anti-obese activity of hexane extract of *Cyperus tubers*.

Anti-tumor/carcinogenic property

In cancer, cells divide uncontrollably and destroy the body tissue. Several herbs and supplements are used as a natural treatment for cancer therapy. Kilani-Jaziri et al. [26] reported that the Ethyl Acetate (EA) and TOF enriched extracts inhibit K562 erythroleukemia cells. The inhibition of K562 cell proliferation might be due to apoptosis induction and activation of Phosphatidylinositol-3-Kinase (PI3K) and Mitogen-Activated Protein Kinases (MAPKs). Ethanol extract of *C. rotundus* showed anti-proliferative activity on human breast carcinoma MDA-MB-231 cells. It was associated with the induction of apoptosis by death receptors and survivin and Bcl-2 activity [70]. Nam et al. [71] revealed that hexane fraction of valencene from *C. rotundus* had therapeutic effects on UV-induced photoaging and decreased the melanin content after UVB irradiation in murine B16F10 melanoma cells. Amentoflavone reduced uterine tumors in rats. This mechanism might be due to higher Bax protein expression, reduced expression of Bcl-2, and progesterone-mediated apoptosis in uterine fibroid cells [72]. The methanolic extract of *C. rotundus* rhizome had

a cytotoxic effect on different cancer cell lines ranging from 4.52 ± 0.57 to 9.85 ± 0.68 $\mu\text{g/mL}$, and its inhibitory effect was shown by migration assay [73]. The n-hexane fraction of an ethanol extract from *C. rotundus* (6-acetoxy cyperene) is an anti-tumor compound that causes caspase-dependent apoptosis to inhibit cell growth in ovarian cancer and endometrial cancer cells [74].

Immunomodulatory and anti-allergic property

Inflammation is the body's defense mechanism to the external noxious physical or chemical stimuli triggered by invading pathogens and is a critical component of various pathological events [55]. Seo et al. [65] reported the anti-inflammatory activity of methanolic extract of *Cyperus tubers* by in vitro murine macrophage cell line RAW 264.7 model challenged with the endotoxin from Gram-negative bacteria, i.e., lipopolysaccharide. The cells showed characteristic features of inflammation, such as the release of nitric oxide by overexpression of iNOS and H_2O_2 radical production. Here the authors demonstrated the protective activity of *Cyperus* extract on endotoxin-induced nitric oxide expression, where 100 μg of the methanolic extract efficiently suppressed iNOS protein and mRNA overexpression and also exhibited anti-oxidative property and suppressed the free radicals suggesting the anti-inflammatory activity of methanolic extract of *Cyperus rotundus*. Dang et al. [75] further reported the anti-inflammatory activity of *Cyperus rotundus* tuber water extract in an in vivo model of carrageenan-induced paw edema and acetic acid-induced peritonitis. Furthermore, 270 mg/kg dose

of the extract showed 9.5% inhibition of the paw edema, and 390 mg/kg dose of extract inhibited the acetic acid-induced peritonitis.

In another study, Jin et al. [76] reported the anti-allergic activity of constituents isolated from *Cyperus*, namely valencene, nootkatone, caryophyllene α -oxide, β -pinene, 1,8-cineole, limonene, and 4-cymene by fractionation on rat basophilic leukemia-1 (RBL-1) cells and in an animal model of delayed-type hypersensitivity. Valencene, nootkatone, and caryophyllene α -oxide showed inhibition of cysteinyl leukotrienes by 60%, 93%, and 99 %, respectively, in calcium ionophore A23187 challenged RBL-1 cells. In another model, RBL-2H3 cells were sensitized by anti-DNP and treated with dinitrophenyl BSA (bovine serum albumin), released β -hexosaminidase by degranulation, which is a biomarker of allergy. *Cyperus* ethanolic extract at 300 μ g/mL inhibited the degranulation by 21%, whereas the sesquiterpene valencene at 100 μ M showed 88% inhibition. In mast cells, Lyn phosphorylation is the critical event in receptor-mediated degranulation. Lyn phosphorylation was measured by western blotting in degranulated RBL-2H3 cells, where 300 μ g/mL CRE and 50 μ M valencene inhibited the Lyn activation. Besides, the anti-allergic activity was evaluated in the hapten-treated mouse delayed hypersensitivity model, where the CRE extract, sesquiterpenes valence, and nootkatone reduced the ear thickness.

Recently, Tsoyi et al. [7] evaluated the anti-inflammatory activity of CRE and its isolated constituents in RAW 264.7 cells and mice model of sepsis. The authors found overexpression of Heme Oxygenase-1 (HO-1) with ethanolic extract, isolated compounds of *Cyperus*, namely nootkatone, valencene, β -selinene, α -cyperone, also downregulated iNOS expression in endotoxin-treated macrophages. The overexpression of HO-1 plays a pivotal role as cytoprotective, anti-apoptotic, and immunomodulator in various stresses. A similar trend was observed in the down-regulation of iNOS expression by nootkatone, valencene, and ethanolic extracts in HO-1-silenced, and LPS-challenged (lipopolysaccharide) cells, suggesting that anti-inflammatory effect is mediated by HO-1 upregulation. Overexpression of cytokine, high mobility group box 1 protein (HMGB1) plays a critical role in sepsis-induced death. In contrast, the downregulation of HMGB1 expression with LPS challenge by HO-1 upregulation was observed with nootkatone, valencene, and ethanolic extracts. Also, it increased the survival rates of Cecal Ligation and Puncture (CLP)-induced sepsis in mice.

Anti-malarial property

Malaria is a life-threatening mosquito-borne blood disease caused by parasites transmitted to people. Traditional medicines have been used as anti-malarial therapy by tribal and folklore medicine. Thebtaranonth et al. [77] explored the anti-malarial activity of *C. rotundus*. Among tested compounds, 10, 12-peroxyvalencene had a maximum effect with an EC₅₀ of 2.33×10^{-6} M and dichloromethane extract with an IC₅₀ between 10 and 50 μ g/mL [78, 79].

Anti-diarrheal property

Diarrhea is a leading cause of malnutrition and death in children under five years old. Traditional medicine is popular for the treatment of acute diarrhea. The anti-diarrheal activity of different fractions (methanolic and petroleum ether) of *C. rotundus* rhizome exhibited significant activity in castor oil-induced diarrhea in mice [80]. Daswani et al. [81] studied the role of decoction of *C. rotundus* against diarrhea and in the absence of antimicrobial activity due to the mechanism of bacterial virulence. Venkatasubramanian et al. [82] reported 46% inhibition of the diarrheal activity of methanolic extract of *C. rotundus*.

Gastroprotective property

Gastroprotective agents protect the stomach and gastric system from various forms of ulcers and gastric tissue damage. Guldur et al. [83] reported the gastroprotective effect of methanolic extract of *Cyperus* rhizomes in a model of an ischemia-reperfusion-induced stomach injury. The administration of the extract effectively restored the anti-oxidant status measured by glutathione and lipid peroxidation products and also protected the microvillus architecture of the stomach with the ischemic model but to a lesser extent in the ischemia-reperfusion model, explaining the potential gastroprotective effect of the extract.

Protein oxidation and protein nitration prevention property

The excess amount of ROS in the presence of \bullet NO or \bullet NO-derived metabolites leads to the formation of nitrating species such as peroxynitrite [20, 84]. The protein oxidation and nitration of tyrosine residues to 3-nitrotyrosine disrupt nitric oxide (\bullet NO) signaling towards pro-oxidant processes and alters the metabolism [85]. Ardestani and Yazdanparast et al. [86] observed that extracts of *C. rotundus* suppresses AGE formation in a fructose-mediated protein glycoxidation model in vitro and inhibit oxidative stress. We also determined that *C. rotundus* inhibits SIN-1 induced protein nitration and

AAPH-induced protein oxidation in both in vitro and ex-vivo models [13, 54] (Figure 6).

Pharmacological Effects of *C. Rotundus* On Human Subjects/Human Cell Lines

Humans consume a wide variety of herbs, spices, and vegetables along with animal and dietary products as part of the diet, which play a key role in maintaining the well-being of an individual. Various herbal and food products are demonstrated to possess medicinal values. In a previous study, *C. rotundus* aqueous extract was shown to have anti-inflammatory activity for conjunctivitis in human subjects [87]. In another study, Appaji et al. [88] investigated the immunoglobulin-enhancing effects of Bala compound, a combination of several plants along with *C. rotundus*. The orally administered Bala compound enhanced immunoglobulin content in infants compared with the multivitamin supplementation group suggesting the potential immunomodulatory activity of the plant extract. Recently, we evaluated the neuroprotective effects of *C. rotundus* against oxidative stress-mediated neuronal damage of SH-SY5Y human neuronal cells [11, 54].

Conclusion and Future Perspectives

The common neurological diseases affecting people across the globe include Alzheimer, Parkinson, epilepsy, ischemia, and its associated neurodegeneration with cognitive decline. *C. rotundus* is a weed, but it is identified to possess various pharmacological properties, including neuro-pharmacological activities. The root part of the plant has been extensively used as a traditional medicine to cure various ailments and used to develop nutraceuticals and pharmacological products. The advancement of scientific methodologies and various in vitro and in vivo preclinical animal models determined the pleiotropic activities of mustaka and its constituents as ethno-medicine. Although various metabolites of *C. rotundus* have been identified, only a few have been thoroughly investigated for the molecular mechanism/mode of action. *C. rotundus* has also been used in combination with other plants as a combinational therapy to treat several diseases. The reported cytotoxic/toxicological effects of *C. rotundus* should be taken into account. However, a little investigation has been carried out on the pharmacokinetic and pharmacodynamic properties of mustaka and its chemical constituents. Furthermore, the full utility of this plant will be more beneficial in the coming years and has great potential for use in the pharmaceutical industry. The phytochemistry of biochemical constituents of

C. rotundus from various researchers across the globe by various analytical techniques demonstrated the genetic diversity of the plant-based on its cultivation in different agro-climatic zones. Furthermore, pharmacokinetics, pharmacodynamics, bioavailability, and pharmacognosy studies are warranted for confirming the phytochemicals responsible for various pharmacological actions and their method and mode of action at the molecular and cellular level and clinical safety in humans.

Ethical Considerations

Compliance with ethical guidelines

There were no ethical considerations to be considered in this research.

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Authors' contributions

All authors equally contributed to preparing this article.

Conflict of interest

The authors declared no conflict of interest.

References

- [1] Pandeya G. Caraka samhita of agnivesa with cakrapanidatta tika. Varanasi: Chaukhamba Sanskrit Sansthan; 1997.
- [2] Umerie S, Ezeuzo H. Physicochemical characterization and utilization of *Cyperus rotundus* starch. *Bioresour Technol.* 2000; 72(2):193-6. [DOI:10.1016/S0960-8524(99)00103-0]
- [3] Eltilib HH, Elgasim EA, Mohamed Ahmed IA. Effect of incorporation of *Cyperus rotundus* L. Rhizome powder on quality attributes of minced beef meat. *J Food Sci Technol.* 2016; 53(9):3446-54. [DOI:10.1007/s13197-016-2317-6] [PMID] [PMCID]
- [4] Singh SP, Raghavendra K, Dash AP. evaluation of hexane extract of tuber of root of *Cyperus rotundus* linn (cyperaceae) for repellency against mosquito vectors. *J Parasitol Res.* 2009; 908085. [DOI:10.1155/2009/908085] [PMID] [PMCID]
- [5] Hu QP, Cao XM, Hao DL, Zhang LL. Chemical composition, antioxidant, dna damage protective, cytotoxic and antibacterial activities of *Cyperus rotundus* rhizomes essential oil against foodborne pathogens. *Sci Rep.* 2017; 7:45231. [DOI:10.1038/srep45231] [PMID] [PMCID]

- [6] Lemaure B, Touché A, Zbinden I, Moulin J, Courtois D, Macé K, et al. Administration of *Cyperus rotundus* tubers extract prevents weight gain in obese Zucker rats. *Phytother Res.* 2007; 21(8):724-30. [DOI:10.1002/ptr.2147] [PMID]
- [7] Tsoyi K, Jang HJ, Lee YS, Kim YM, Kim HJ, Seo HG, et al. (+)-Nootkatone and (+)-valencene from rhizomes of *Cyperus rotundus* increase survival rates in septic mice due to heme oxygenase-1 induction. *J Ethnopharmacol.* 2011; 137(3):1311-7. [DOI:10.1016/j.jep.2011.07.062] [PMID]
- [8] Sunil AG, Kesavanarayanan KS, Kalaivani P, Sathiyaa S, Ranju V, Priya RJ, et al. Total oligomeric flavonoids of *Cyperus rotundus* ameliorates neurological deficits, excitotoxicity and behavioral alterations induced by cerebral ischemic-reperfusion injury in rats. *Brain Res Bull.* 2011; 84(6):394-405. [DOI:10.1016/j.brainresbull.2011.01.008] [PMID]
- [9] Kumar KH, Razack S, Nallamuthu I, Khanum F. Phytochemical analysis and biological properties of *Cyperus rotundus* L. *Ind Crops Prod.* 2014;52:815-26. [DOI:10.1016/j.indcrop.2013.11.040]
- [10] Kandikattu HK, Deep SN, Razack S, Amruta N, Prasad D, Khanum F. Hypoxia induced cognitive impairment modulating activity of *Cyperus rotundus*. *Physiol Behav.* 2017; 175:56-65. [DOI:10.1016/j.physbeh.2017.03.035] [PMID]
- [11] Kumar KH, Khanum F. Hydroalcoholic extract of *Cyperus rotundus* ameliorates H₂O₂-induced human neuronal cell damage via its anti-oxidative and anti-apoptotic machinery. *Cell Mol Neurobiol.* 2013; 33(1):5-17. [DOI:10.1007/s10571-012-9865-8] [PMID]
- [12] Venuprasad MP, Hemanth Kumar K, Khanum F. Neuroprotective effects of hydroalcoholic extract of *ocimum sanctum* against H₂O₂ induced neuronal cell damage in SH-SY5Y cells via its antioxidative defence mechanism. *Neurochem Res.* 2013; 38(10):2190-200. [DOI:10.1007/s11064-013-1128-7] [PMID]
- [13] Kandikattu HK, Rachitha P, Krupashree K, Jayashree GV, Abhishek V, Khanum F. LC-ESI-MS/MS analysis of total oligomeric flavonoid fraction of *Cyperus rotundus* and its antioxidant, macromolecule damage protective and antihemolytic effects. *Pathophysiology.* 2015; 22(4):165-73. [DOI:10.1016/j.pathophys.2015.07.001] [PMID]
- [14] Pirzada AM, Ali HH, Naeem M, Latif M, Bukhari AH, Tanveer A. *Cyperus rotundus* L: Traditional uses, phytochemistry, and pharmacological activities. *J Ethnopharmacol.* 2015; 174:540-60. [DOI:10.1016/j.jep.2015.08.012] [PMID]
- [15] Lansdown RV, Juffe Bignoli D, Beentje HJ. *Cyperus rotundus*. The IUCN Red List of Threatened Species. 2017; 2017: e.T158183A84284983.
- [16] Qasim M, Abideen Z, Adnan MY, Ansari R, Gul B, Khan M. Traditional ethnobotanical uses of medicinal plants from coastal areas. *J coast life Med* 2014;2(1):22-30. https://www.halophyte.org/pdfs/drkhan_pdfs/191.pdf
- [17] Hort A. Enquiry into plants and minor works on odours and weather signs, Volume 2. Theophrastus: the University of Michigan. 1916. https://www.google.com/books/edition/Enquiry_Into_Plants_and_Minor_Works_on_O/vJLAAAAMAAJ?hl=en&gbpv=0
- [18] David WH, Jason AF. Purple nutsedge, *Cyperus rotundus* L. Florida (U.S.A): Institute of Food and Agricultural Sciences. University of Florida, 2012.
- [19] Holm LG, Plucknett DL, Pancho JV, Herberger JP. The world's worst weeds. Distribution and biology: University press of Hawaii; 1977. <https://www.cabi.org/isc/abstract/19782319336>
- [20] Jayashree G, Kumar KH, Krupashree K, Rachitha P, Khanum F. LC-ESI-MS/MS analysis of *Asparagus racemosus* willd. roots and its protective effects against t-BHP induced oxidative stress in rats. *Ind Crops Prod.* 2015; 78:102-9. [DOI:10.1016/j.indcrop.2015.10.024]
- [21] Razack S, Kumar KH, Nallamuthu I, Naika M, Khanum F. Antioxidant, biomolecule oxidation protective activities of *nardostachys jatamansi* dc and its phytochemical analysis by RP-HPLC and GC-MS. *Antioxidants (Basel).* 2015; 4(1):185-203. [DOI:10.3390/antiox4010185] [PMID] [PMCID]
- [22] Sonwa MM, König WA. Chemical study of the essential oil of *Cyperus rotundus*. *Phytochemistry.* 2001; 58(5):799-810. [DOI:10.1016/s0031-9422(01)00301-6] [PMID]
- [23] Jeong SJ, Miyamoto T, Inagaki M, Kim YC, Higuchi R. Rotundines A-C, three novel sesquiterpene alkaloids from *Cyperus rotundus*. *J Nat Prod.* 2000; 63(5):673-5. [DOI:10.1021/np990588r] [PMID]
- [24] Chen Y, Zhao YY, Wang XY, Liu JT, Huang LQ, Peng CS. GC-MS analysis and analgesic activity of essential oil from fresh rhizoma of *Cyperus rotundus*. *Zhong Yao Cai.* 2011; 34(8):1225-9. [PMID]
- [25] Jayashree G, Rachitha P, Krupashree K, Kumar K, Khanum F. Phytochemical analysis of methanolic extract of roots of *Asparagus racemosus* (Shatavari). *Int J Pharma Bio Sci.* 2013; 4(4):250-4. <https://www.ijpbs.net/abstract.php?article=MjcxMA==>
- [26] Kilani-Jaziri S, Neffati A, Limem I, Boubaker J, Skandrani I, Sghair MB, et al. Relationship correlation of antioxidant and antiproliferative capacity of *Cyperus rotundus* products towards K562 erythroleukemia cells. *Chem Biol Interact.* 2009; 181(1):85-94. [DOI:10.1016/j.cbi.2009.04.014] [PMID]
- [27] Kandikattu HK, Venuprasad MP, Pal A, Khanum F. Phytochemical analysis and exercise enhancing effects of hydroalcoholic extract of *celastrus paniculatus* willd. *Ind Crops Prod.* 2014 ;55:217-24. [DOI:10.1016/j.indcrop.2014.01.043]
- [28] Venuprasad MP, Kandikattu HK, Razack S, Amruta N, Khanum F. Chemical composition of *ocimum sanctum* by LC-ESI-MS/MS analysis and its protective effects against smoke induced lung and neuronal tissue damage in rats. *Biomed Pharmacother.* 2017; 91:1-12. [DOI:10.1016/j.biopha.2017.04.011] [PMID]
- [29] Cheignon C, Tomas M, Bonnefont-Rousselot D, Faller P, Hureau C, Collin F. Oxidative stress and the amyloid beta peptide in Alzheimer's disease. *Redox Biol.* 2018; 14:450-64. [DOI:10.1016/j.redox.2017.10.014] [PMID] [PMCID]
- [30] Mehdizadeh M, Hashem Dabaghian F, Shojaee A, Molavi N, Taslimi Z, Shabani R, et al. [Protective effects of *Cyperus rotundus* extract on amyloid β -peptide (1-40)-induced memory impairment in male rats: A behavioral study (Persian)]. *Basic Clin Neurosci.* 2017; 8(3):249-54. [DOI:10.18869/nirp.bcn.8.3.249] [PMID] [PMCID]
- [31] Azimi A, Ghaffari SM, Riazi GH, Arab SS, Tavakol MM, Pooyan S. A-cyperone of *Cyperus rotundus* is an effective candidate for reduction of inflammation by destabilization of microtubule fibers in brain. *J Ethnopharmacol.* 2016; 194:219-27. [DOI:10.1016/j.jep.2016.06.058] [PMID]

- [32] Lee CH, Hwang DS, Kim HG, Oh H, Park H, Cho JH, et al. Protective effect of cyperi rhizoma against 6-hydroxydopamine-induced neuronal damage. *J Med Food*. 2010; 13(3):564-71. [DOI:10.1089/jmf.2009.1252] [PMID]
- [33] Venuprasad MP, Kandikattu HK, Razack S, Khanum F. Phytochemical analysis of ocimum gratissimum by LC-ESI-MS/MS and its antioxidant and anxiolytic effects. *S Afr J Bot*. 2014; 92:151-8. [DOI:10.1016/j.sajb.2014.02.010]
- [34] Razack S, Kandikattu HK, Venuprasad MP, Amruta N, Khanum F, Chuttani K, et al. Anxiolytic actions of nardostachys jatamansi via GABA benzodiazepine channel complex mechanism and its biodistribution studies. *Metab Brain Dis*. 2018; 33(5):1533-49. [DOI:10.1007/s11011-018-0261-z] [PMID]
- [35] Rudolph U, Knoflach F. Beyond classical benzodiazepines: Novel therapeutic potential of GABAA receptor subtypes. *Nat Rev Drug Discov*. 2011; 10(9):685-97. [DOI:10.1038/nrd3502] [PMID] [PMCID]
- [36] Wei XH, Cheng XM, Shen JS, Wang ZT. Antidepressant effect of yueju-wan ethanol extract and its fractions in mice models of despair. *J Ethnopharmacol*. 2008; 117(2):339-44. [DOI:10.1016/j.jep.2008.02.004] [PMID]
- [37] Pal D, Dutta S, Sarkar A. Evaluation of CNS activities of ethanol extract of roots and rhizomes of *Cyperus rotundus* in mice. *Acta Pol Pharm*. 2009; 66(5):535-41. [PMID]
- [38] Khalili M, Kiasalari Z, Roghani M, Azizi Y. Anticonvulsant and antioxidant effect of hydro-alcoholic extract of *Cyperus rotundus* rhizome on pentylenetetrazole-induced kindling model in male mice. *Journal of Medicinal Plants Research*. 2011; 5(7):1140-6. [DOI:10.5897/JMPR.9000137]
- [39] Shivakumar SI, Suresh HM, Hallikeri CS, Hatapakki BC, Handiganur JS, Kuber S, et al. Anticonvulsant effect of *Cyperus rotundus* Linn rhizomes in rats. *J Nat Remedies*. 2009; 9(2):192-6. [DOI:10.18311/jnr/2009/239]
- [40] Ahmad M, Mahayrookh M, Rehman AB, Jahan N. Analgesic, antimicrobial and cytotoxic effect of *Cyperus rotundus* ethanol extract. *Pak J Pharmacol*. 2012; 29(2):7-13. <https://citeseerx.ist.psu.edu/viewdoc/download?doi=10.1.1.1038.568&rep=rep1&type=pdf>
- [41] Pal DK. Determination of brain biogenic amines in *Cynodon dactylon* pers. And *Cyperus rotundus* L. treated mice. *Int J Pharm Pharm Sci*. 2009; 1(1):190-7. <https://innovareacademics.in/journal/ij/Vol%201%20Issue%201.htm>
- [42] Mandal SC, Dhara AK, Maiti BC. Studies on psychopharmacological activity of *andrographis paniculata* extract. *Phytother Res*. 2001; 15(3):253-6. [DOI:10.1002/ptr.704] [PMID]
- [43] Imam MZ, Sumi CD. Evaluation of antinociceptive activity of hydromethanol extract of *Cyperus rotundus* in mice. *BMC Complement Altern Med*. 2014; 14:83. [DOI:10.1186/1472-6882-14-83] [PMID] [PMCID]
- [44] Rachitha P, Krupashree K, Jayashree GV, Kandikattu HK, Amruta N, Gopalan N, et al. Chemical composition, antioxidant potential, macromolecule damage and neuroprotective activity of *convolvulus pluricaulis*. *J Tradit Complement Med*. 2018; 8(4):483-96. [DOI:10.1016/j.jtcme.2017.11.002] [PMID] [PMCID]
- [45] Pandareesh MD, Kandikattu HK, Razack S, Amruta N, Choudhari R, Vikram A, et al. Nutrition and nutraceuticals in neuroinflammatory and brain metabolic stress: Implications for neurodegenerative disorders. *CNS Neurol Disord Drug Targets*. 2018; 17(9):680-8. [DOI:10.2174/1871527317666180625104753] [PMID]
- [46] Ha JH, Lee KY, Choi HC, Cho J, Kang BS, Lim JC, et al. Modulation of radioligand binding to the GABA(A)-benzodiazepine receptor complex by a new component from *Cyperus rotundus*. *Biol Pharm Bull*. 2002; 25(1):128-30. [DOI:10.1248/bpb.25.128] [PMID]
- [47] Lee MS, Lee J, Kwon DY, Kim MS. Ondamtanggamibang protects neurons from oxidative stress with induction of heme oxygenase-1. *J Ethnopharmacol*. 2006; 108(2):294-8. [DOI:10.1016/j.jep.2006.05.012] [PMID]
- [48] Dabaghian FH, Hashemi M, Entezari M, Movassaghi S, Goushegir SA, Kalantari S, et al. [Effect of *Cyperus rotundus* on ischemia-induced brain damage and memory dysfunction in rats (Persian)]. *Iran J Basic Med Sci*. 2015; 18(2):199-204. [PMID] [PMCID]
- [49] Satalangka C, Wattanathorn J. Neuroprotective and cognitive-enhancing effects of the combined extract of *Cyperus rotundus* and *Zingiber officinale*. *BMC Complement Altern Med*. 2017; 17(1):135. [DOI:10.1186/s12906-017-1632-4] [PMID] [PMCID]
- [50] Jebasingh D, Devavaram Jackson D, Venkataraman S, Adeghate E, Starling Emerald B. The protective effects of *Cyperus rotundus* on behavior and cognitive function in a rat model of hypoxia injury. *Pharm Biol*. 2014; 52(12):1558-69. [DOI:10.3109/13880209.2014.908395] [PMID]
- [51] Sharma R, Gupta R. *Cyperus rotundus* extract inhibits acetylcholinesterase activity from animal and plants as well as inhibits germination and seedling growth in wheat and tomato. *Life Sci*. 2007; 80(24-25):2389-92. [DOI:10.1016/j.lfs.2007.01.060] [PMID]
- [52] Krupashree K, Kumar KH, Rachitha P, Jayashree GV, Khanum F. Chemical composition, antioxidant and macromolecule damage protective effects of *Picrorhiza kurroa* Royle ex Benth. *S Afr J Bot*. 2014; 94:249-54. [DOI:10.1016/j.sajb.2014.07.001]
- [53] Pal A, Kumar KH, Bhushan B, Saharan V. *Ashwagandha* root extract inhibits acetylcholine esterase, protein modification and ameliorates H₂O₂-induced oxidative stress in rat lymphocytes. *Pharmacogn J*. 2017;9(3). [DOI:10.5530/pj.2017.3.52] <https://www.phcoj.com/article/320>
- [54] Hemanth Kumar K, Tamatam A, Pal A, Khanum F. Neuroprotective effects of *Cyperus rotundus* on SIN-1 induced nitric oxide generation and protein nitration: Ameliorative effect against apoptosis mediated neuronal cell damage. *Neurotoxicology*. 2013; 34:150-9. [DOI:10.1016/j.neuro.2012.11.002] [PMID]
- [55] Kandikattu HK, Rachitha P, Jayashree GV, Krupashree K, Sukhith M, Majid A, et al. Anti-inflammatory and antioxidant effects of cardamom (*elettaria repens* (Sonn) baill) and its phytochemical analysis by 4D GCXGC TOF-MS. *Biomed Pharmacother*. 2017; 91:191-201. [DOI:10.1016/j.biopha.2017.04.049] [PMID]
- [56] Reddy T, Moulali A, Anjaneyulu E, Ramgopal M, Kumar H, Lokanatha O, et al. Antimicrobial screening of the plant extracts of *Cardiospermum halicacabum* L. against selected microbes. *Ethnobot Leaflets* 2010;14:911-9. https://www.researchgate.net/230892538_Antimicro_microbes
- [57] Sini S, Malathy NS. Antimicrobial properties of roots of medicinal plants. *Anc Sci Life*. 2005; 25(2):62-5. [PMID] [PMCID]

- [58] Jabier MS, Wagi RI, Hussain HAA-K. Extraction, identification and antibacterial activity of *Cyperus* oil from Iraqi *C. rotundus*. *Eng Technol J* 2008; 26(10):1156-63. <https://www.iasj.net/iasj/article/26847>
- [59] Kilani S, Ben Sghaier M, Limem I, Bouhleb I, Boubaker J, Bhouiri W, et al. In vitro evaluation of antibacterial, antioxidant, cytotoxic and apoptotic activities of the tubers infusion and extracts of *Cyperus rotundus*. *Bioresour Technol*. 2008; 99(18):9004-8. [DOI:10.1016/j.biortech.2008.04.066] [PMID]
- [60] Kilani-Jaziri S, Bhouiri W, Skandrani I, Limem I, Chekir-Ghedira L, Ghedira K. Phytochemical, antimicrobial, antioxidant and antigenotoxic potentials of *Cyperus rotundus* extracts. *S Afr J Bot*. 2011; 77(3):767-76. [DOI:10.1016/j.sajb.2011.03.015]
- [61] Tambekar DH, Khante BS, Chandak BR, Titare AS, Borkar SS, Aghadte SN. Screening of antibacterial potentials of some medicinal plants from Melghat forest in India. *Afr J Tradit Complement Altern Med*. 2009; 6(3):228-32. [DOI:10.4314/ajtcam.v6i3.57158] [PMID] [PMCID]
- [62] Kilani S, Ben Ammar R, Bouhleb I, Abdelwahed A, Hayder N, Mahmoud A, et al. Investigation of extracts from (Tunisian) *Cyperus rotundus* as antimutagens and radical scavengers. *Environ Toxicol Pharmacol*. 2005; 20(3):478-84. [DOI:10.1016/j.etap.2005.05.012] [PMID]
- [63] Nagulendran K, Velavan S, Mahesh R, Begum VH. *In vitro* antioxidant activity and total polyphenolic content of *Cyperus rotundus* rhizomes. *J Chem*. 2007; 4(3):440-9. [DOI:10.1155/2007/903496]
- [64] Kilani S, Ledauphin J, Bouhleb I, Ben Sghaier M, Boubaker J, Skandrani I, et al. Comparative study of *Cyperus rotundus* essential oil by a modified GC/MS analysis method. Evaluation of its antioxidant, cytotoxic, and apoptotic effects. *Chem Biodivers*. 2008; 5(5):729-42. [DOI:10.1002/cbdv.200890069] [PMID]
- [65] Seo WG, Pae HO, Oh GS, Chai KY, Kwon TO, Yun YG, et al. Inhibitory effects of methanol extract of *Cyperus rotundus* rhizomes on nitric oxide and superoxide productions by murine macrophage cell line, RAW 264.7 cells. *J Ethnopharmacol*. 2001; 76(1):59-64. [DOI:10.1016/s0378-8741(01)00221-5] [PMID]
- [66] Kakarla L, Katragadda SB, Tiwari AK, Kotamraju KS, Madhusudana K, Kumar DA, et al. Free radical scavenging, α -glucosidase inhibitory and anti-inflammatory constituents from Indian sedges, *Cyperus scariosus* R.Br and *Cyperus rotundus* L. *Pharmacogn Mag*. 2016; 12(Suppl 4):S488-S96. [DOI:10.4103/0973-1296.191467] [PMID] [PMCID]
- [67] Lydia J, Sudarsanam A. Docking of a *Cyperus rotundus* compound '15-Hydroxy-4-oxo-10-pentadecynoic acid lactone' with antidiabetic drug targets: a comparative study. *Int J Fund Appl Sci*. 2014; 3(2):17-21. <https://www.semanticscholar.org/paper/Docking-of-a-Odaeb725cd38>
- [68] Singh P, Khosa RL, Mishra G, Jha KK. Antidiabetic activity of ethanolic extract of *Cyperus rotundus* rhizomes in streptozotocin-induced diabetic mice. *J Pharm Bioallied Sci*. 2015; 7(4):289-92. [DOI:10.4103/0975-7406.168028] [PMID] [PMCID]
- [69] Bambhole VD. Effect of some medicinal plant preparations of adipose tissue metabolism. *Anc Sci Life*. 1988; 8(2):117-24. [PMID] [PMCID]
- [70] Park SE, Shin WT, Park C, Hong SH, Kim GY, Kim SO, et al. Induction of apoptosis in MDA-MB-231 human breast carcinoma cells with an ethanol extract of *Cyperus rotundus* L. by activating caspases. *Oncol Rep*. 2014; 32(6):2461-70. [DOI:10.3892/or.2014.3507] [PMID]
- [71] Nam JH, Nam DY, Lee DU. Valencene from the rhizomes of *Cyperus rotundus* inhibits skin photoaging-related Ion channels and UV-Induced Melanogenesis in B16F10 Melanoma cells. *J Nat Prod*. 2016; 79(4):1091-6. [DOI:10.1021/acs.jnatprod.5b01127] [PMID]
- [72] Ying J, Bing X. Chemical constituents of *Cyperus rotundus* L. And their inhibitory effects on uterine fibroids. *Afr Health Sci*. 2016; 16(4):1000-6. [DOI:10.4314/ahs.v16i4.16] [PMID] [PMCID]
- [73] Mannarreddy P, Denis M, Munireddy D, Pandurangan R, Thangavelu KP, Venkatesan K. Cytotoxic effect of *Cyperus rotundus* rhizome extract on human cancer cell lines. *Biomed Pharmacother*. 2017; 95:1375-87. [DOI:10.1016/j.biopha.2017.09.051] [PMID]
- [74] Ahn JH, Lee TW, Kim KH, Byun H, Ryu B, Lee KT, et al. 6-Acetoxy cyperene, a patchoulane-type sesquiterpene isolated from *Cyperus rotundus* rhizomes induces caspase-dependent apoptosis in human ovarian cancer cells. *Phytother Res*. 2015; 29(9):1330-8. [DOI:10.1002/ptr.5385] [PMID]
- [75] Dang GK, Parekar RR, Kamat SK, Scindia AM, Rege NN. Antiinflammatory activity of *phyllanthus emblica*, *plumbago zeylanica* and *Cyperus rotundus* in acute models of inflammation. *Phytother Res*. 2011; 25(6):904-8. [DOI:10.1002/ptr.3345] [PMID]
- [76] Jin JH, Lee DU, Kim YS, Kim HP. Anti-allergic activity of sesquiterpenes from the rhizomes of *Cyperus rotundus*. *Arch Pharm Res*. 2011; 34(2):223-8. [DOI:10.1007/s12272-011-0207-z] [PMID]
- [77] Thebtaranonth C, Thebtaranonth Y, Wanauppathamkul S, Yuthavong Y. Antimalarial sesquiterpenes from tubers of *Cyperus rotundus*: Structure of 10,12-peroxy-calamenene, a sesquiterpene endoperoxide. *Phytochemistry*. 1995; 40(1):125-8. [DOI:10.1016/0031-9422(95)00260-e] [PMID]
- [78] Weenen H, Nkonya MH, Bray DH, Mwasumbi LB, Kinabo LS, Kilimali VA. Antimalarial activity of Tanzanian medicinal plants. *Planta Med*. 1990; 56(4):368-70. [DOI:10.1055/s-2006-960984] [PMID]
- [79] Kamala A, Middha SK, Karigar CS. Plants in traditional medicine with special reference to *Cyperus rotundus* L.: A review. *3 Biotech*. 2018; 8(7):309. [DOI:10.1007/s13205-018-1328-6] [PMID] [PMCID]
- [80] Uddin SJ, Mondal K, Shilpi JA, Rahman MT. Antidiarrhoeal activity of *Cyperus rotundus*. *Fitoterapia*. 2006; 77(2):134-6. [DOI:10.1016/j.fitote.2004.11.011] [PMID]
- [81] Daswani PG, Brijesh S, Tetali P, Birdi TJ. Studies on the activity of *Cyperus rotundus* Linn. tubers against infectious diarrhea. *Indian J Pharmacol*. 2011; 43(3):340-4. [DOI:10.4103/0253-7613.81502] [PMID] [PMCID]
- [82] Venkatasubramanian P, Kumar SK, Nair VS. *Cyperus rotundus*, a substitute for *aconitum heterophyllum*: Studies on the ayurvedic concept of *abhava pratinidhi dravya* (drug substitution). *J Ayurveda Integr Med*. 2010; 1(1):33-9. [DOI:10.4103/0975-9476.59825] [PMID] [PMCID]
- [83] Muhammet EG, OZgönül A, Kilic IH, Söğüt O, Ozaşlan M, Bitiren M, et al. Gastroprotective effect Of *Cyperus rotundus*

- extract against gastric mucosal injury induced by ischemia and reperfusion in rats. *International Journal of Pharmacology*. 2010; 6:(2):104-10. [DOI:10.3923/ijp.2010.104.110]
- [84] Jayashree GV, Rachitha P, Krupashree K, Kumar KH, Khanum F. Antioxidant and DNA damage protective effects of *Asparagus racemosus* in human colon and mice muscle cells. *Pharmacognosy Journal*. 2015; 7(3):182-90. <https://www.phcogj.com/article/1442>
- [85] Radi R. Protein tyrosine nitration: Biochemical mechanisms and structural basis of functional effects. *Acc Chem Res*. 2013; 46(2):550-9. [DOI:10.1021/ar300234c][PMID] [PMCID]
- [86] Ardestani A, Yazdanparast R. [Cyperus rotundus suppresses AGE formation and protein oxidation in a model of fructose-mediated protein glycoxidation (Persian)]. *Int J Biol Macromol*. 2007; 41(5):572-8. [DOI:10.1016/j.jbiomac.2007.07.014] [PMID]
- [87] Saxena R, Punhami PT, Garg K, Singh N, Kohli R. Preliminary report on the anti-inflammatory activity of *Cyperus rotundus* in conjunctivitis (in human subjects). *Indian J Pharm* 1971; 3:9.
- [88] Appaji RR, Sharma RD, Katiyar GP, Sai PA. Clinical study of the immunoglobulin enhancing effect of "bala compound" on Infants. *Anc Sci Life*. 2009; 28(3):18-22. [PMID] [PMCID]
- [89] Quintans-Júnior L, Moreira JC, Pasquali MA, Rabie SM, Pires AS, Schröder R, et al. Antinociceptive activity and redox profile of the monoterpenes (+)-camphene, p-cymene, and geranyl acetate in experimental models. *ISRN Toxicol*. 2013; 2013:459530. [DOI:10.1155/2013/459530][PMID] [PMCID]
- [90] Djilani A, Dicko A. The therapeutic benefits of essential oils. *Nutrition, Well-being and Health*. 2012; 7:155-79. <https://www.intechopen.com/chapters/29979>
- [91] Lulekal E, Rondevaldova J, Bernaskova E, Cepkova J, Asfaw Z, Kelbessa E, et al. Antimicrobial activity of traditional medicinal plants from ankober district, north shewa zone, amhara region, ethiopia. *Pharm Biol*. 2014; 52(5):614-20. [DOI:10.3109/13880209.2013.858362] [PMID]
- [92] Arunkumar R, Nair SA, Rameshkumar KB, Subramoniam A. The essential oil constituents of *Zornia diphylla* (L.) Pers, and anti-inflammatory and antimicrobial activities of the oil. *Rec Nat Prod*. 2014; 8(4):385-93. <https://www.acgpubs.org/article/records-of-natural-products/2014/4-october-1>
- [93] Rivas da Silva AC, Lopes PM, Barros de Azevedo MM, Costa DC, Alviano CS, Alviano DS. Biological activities of α -pinene and β -pinene enantiomers. *Molecules*. 2012; 17(6):6305-16. [DOI:10.3390/molecules17066305] [PMID] [PMCID]
- [94] Rufino AT, Ribeiro M, Judas F, Salgueiro L, Lopes MC, Cavaleiro C, et al. Anti-inflammatory and chondroprotective activity of (+)- α -pinene: Structural and enantiomeric selectivity. *J Nat Prod*. 2014; 77(2):264-9. [DOI:10.1021/np400828x] [PMID]
- [95] Santos FA, Rao VS. Antiinflammatory and antinociceptive effects of 1, 8-cineole a terpenoid oxide present in many plant essential oils. *Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives*. 2000; 14(4):240-4. [DOI:10.1002/1099-1573(200006)14:4<3C240::AID-PTR573%3E3.0.CO;2-X]
- [96] Asanova ZK, Suleimenov E, Atazhanova G, Dembitskii A, Pak R, Dar A, et al. Biological activity of 1, 8-cineole from levant wormwood. *Pharm Chem J* 2003; 37(1):28-30. [DOI:10.1023/A:1023699012354]
- [97] Nóbrega FF, Salvadori MG, Masson CJ, Mello CF, Nascimento TS, Leal-Cardoso JH, et al. Monoterpenoid terpinen-4-ol exhibits anticonvulsant activity in behavioural and electrophysiological studies. *Oxid Med Cell Longev*. 2014; 2014:703848. [DOI:10.1155/2014/703848] [PMID] [PMCID]
- [98] de Oliveira MG, Marques RB, de Santana MF, Santos AB, Brito FA, Barreto EO, et al. A-terpineol reduces mechanical hypernociception and inflammatory response. *Basic Clin Pharmacol Toxicol*. 2012; 111(2):120-5. [DOI:10.1111/j.1742-7843.2012.00875.x] [PMID]
- [99] Baser KH. Biological and pharmacological activities of carvacrol and carvacrol bearing essential oils. *Curr Pharm Des*. 2008; 14(29):3106-19. [DOI:10.2174/138161208786404227][PMID]
- [100] Cherneva E, Pavlovic V, Smelcerovic A, Yancheva D. The effect of camphor and borneol on rat thymocyte viability and oxidative stress. *Molecules*. 2012; 17(9):10258-66. [DOI:10.3390/molecules170910258] [PMID] [PMCID]
- [101] Kaufmann D, Dogra AK, Wink M. Myrtenal inhibits acetylcholinesterase, a known Alzheimer target. *J Pharm Pharmacol*. 2011; 63(10):1368-71. [DOI:10.1111/j.2042-7158.2011.01344.x] [PMID]
- [102] Aydin E, Türkez H, Taşdemir S. Anticancer and antioxidant properties of terpinolene in rat brain cells. *Arh Hig Rada Toksikol*. 2013; 64(3):415-24. [DOI:10.2478/10004-1254-64-2013-2365] [PMID]
- [103] Riella KR, Marinho RR, Santos JS, Pereira-Filho RN, Cardoso JC, Albuquerque-Junior RL, et al. Anti-inflammatory and cicatrizing activities of thymol, a monoterpene of the essential oil from *Lippia gracilis*, in rodents. *J Ethnopharmacol*. 2012; 143(2):656-63. [DOI:10.1016/j.jep.2012.07.028] [PMID]
- [104] Javan AJ, Javan MJ. [Electronic structure of some thymol derivatives correlated with the radical scavenging activity: Theoretical study (Persian)]. *Food Chem*. 2014; 165:451-9. [DOI:10.1016/j.foodchem.2014.05.073] [PMID]
- [105] Leandro LM, Vargas Fde S, Barbosa PC, Neves JK, da Silva JA, da Veiga-Junior VF. Chemistry and biological activities of terpenoids from copaiba (*Copaifera* spp.) oleoresins. *Molecules*. 2012; 17(4):3866-89. [DOI:10.3390/molecules17043866] [PMID] [PMCID]
- [106] Ananth DA, Sivasudha T, Rameshkumar A, Jeyadevi R, Aseervatham SB. Chemical constituents, in vitro antioxidant and antimicrobial potential of *Caryota urens* L. *Free Radicals and Antioxidants*. 2013; 3(2):107-12. [DOI:10.1016/j.fra.2013.05.008]
- [107] Jung SH, Kim SJ, Jun BG, Lee KT, Hong SP, Oh MS, et al. A-cyperone, isolated from the rhizomes of *Cyperus rotundus*, inhibits LPS-induced COX-2 expression and PGE2 production through the negative regulation of NF κ B signalling in RAW 264.7 cells. *J Ethnopharmacol*. 2013; 147(1):208-14. [DOI:10.1016/j.jep.2013.02.034] [PMID]
- [108] Ding L, Pfoh R, Rühl S, Qin S, Laatsch H. T-muurolol sesquiterpenes from the marine streptomyces sp. M491 and revision of the configuration of previously reported amorphanes. *J Nat Prod*. 2009; 72(1):99-101. [DOI:10.1021/np8006843] [PMID]

- [109] Mevy J, Bessiere J, Dherbomez M, Millogo J, Viano J. Chemical composition and some biological activities of the volatile oils of a chemotype of *Lippia chevalieri* moldenke. *Food Chem.* 2007; 101(2):682-5. [DOI:10.1016/j.foodchem.2006.01.052]
- [110] Rukungu GM, Muregi FW, Omar SA, Gathirwa JW, Muthaura CN, Peter MG, et al. Anti-plasmodial activity of the extracts and two sesquiterpenes from *Cyperus articulatus*. *Fitoterapia.* 2008; 79(3):188-90. [DOI:10.1016/j.fitote.2007.11.010][PMID]
- [111] Ghannadi A, Rabbani M, Ghaemmaghami L, Malekian N. Phytochemical screening and essential oil analysis of one of the Persian sedges; *Cyperus rotundus* L. *Int J Pharm Sci Res.* 2012; 3(2):424-27. [DOI:10.13040/IJPSR.0975-8232.3(2).424-27]
- [112] Solís C, Becerra J, Flores C, Robledo J, Silva M. Antibacterial and antifungal terpenes from pilgerodendron uviferum (D. Don) florin. *J Chil Chem Soc.* 2004; 49(2):157-61. [DOI:10.4067/S0717-97072004000200010]
- [113] Claeson P, Andersson R, Samuelsson G. T-cadinol: A pharmacologically active constituent of scented myrrh: Introductory pharmacological characterization and high field 1H- and 13C-NMR data. *Planta Med.* 1991; 57(4):352-6. [DOI:10.1055/s-2006-960116] [PMID]
- [114] Pramod K, Ansari SH, Ali J. Eugenol: A natural compound with versatile pharmacological actions. *Nat Prod Commun.* 2010; 5(12):1999-2006. [PMID]
- [115] Hari Babu L, Perumal S, Balasubramanian MP. Myrtenal attenuates diethylnitrosamine-induced hepatocellular carcinoma in rats by stabilizing intrinsic antioxidants and modulating apoptotic and anti-apoptotic cascades. *Cell Oncol (Dordr).* 2012; 35(4):269-83. [DOI:10.1007/s13402-012-0086-4] [PMID]
- [116] Mesa-Arango AC, Montiel-Ramos J, Zapata B, Durán C, Betancur-Galvis L, Stashenko E. Citral and carvone chemotypes from the essential oils of Colombian *Lippia alba* (Mill.) N.E. Brown: Composition, cytotoxicity and antifungal activity. *Mem Inst Oswaldo Cruz.* 2009; 104(6):878-84. [DOI:10.1590/s0074-02762009000600010] [PMID]
- [117] Gonçalves JC, Silveira AL, de Souza HD, Nery AA, Prado VF, Prado MA, et al. The monoterpene (-)-carvone: A novel agonist of TRPV1 channels. *Cytometry A.* 2013; 83(2):212-9. [DOI:10.1002/cyto.a.22236] [PMID]
- [118] Dong Y, McCullough KJ, Wittlin S, Chollet J, Vennerstrom JL. The structure and antimalarial activity of dispiro-1,2,4,5-tetraoxanes derived from (+)-dihydrocarvone. *Bioorg Med Chem Lett.* 2010; 20(22):6359-61. [DOI:10.1016/j.bmcl.2010.09.113] [PMID]
- [119] Turkez H, Togar B, Tatar A, Geyikoglu F, Hacimuftuoglu A. Cytotoxic and cytogenetic effects of α -copaene on rat neuron and N2a neuroblastoma cell lines. *Biologia.* 2014; 69(7): 936-942. [DOI:10.2478/s11756-014-0393-5]
- [120] Mulyaningsih S, Sporer F, Zimmermann S, Reichling J, Wink M. Synergistic properties of the terpenoids aromadendrene and 1,8-cineole from the essential oil of *Eucalyptus globulus* against antibiotic-susceptible and antibiotic-resistant pathogens. *Phytomedicine.* 2010; 17(13):1061-6. [DOI:10.1016/j.phymed.2010.06.018] [PMID]
- [121] Dong Q, Yin T, Wang G. Potential applications of beta-ell-emene in anti-atherosclerosis and anti-restenosis. *Sheng Wu Yi Xue Gong Cheng Xue Za Zhi.* 2013; 30(3):656-60. [PMID]
- [122] Govinden-Soulange J, Magan N, Gurib-Fakim A, Gauvin A, Smadja J, Kodja H. Chemical composition and in vitro antimicrobial activities of the essential oils from endemic *Psiadia* species growing in mauritius. *Biol Pharm Bull.* 2004; 27(11):1814-8. [DOI:10.1248/bpb.27.1814] [PMID]
- [123] Legault J, Pichette A. Potentiating effect of beta-caryophyllene on anticancer activity of alpha-humulene, isocaryophyllene and paclitaxel. *J Pharm Pharmacol.* 2007; 59(12):1643-7. [DOI:10.1211/jpp.59.12.0005] [PMID]
- [124] Rogerio AP, Andrade EL, Leite DF, Figueiredo CP, Calixto JB. Preventive and therapeutic anti-inflammatory properties of the sesquiterpene alpha-humulene in experimental airways allergic inflammation. *Br J Pharmacol.* 2009; 158(4):1074-87. [DOI:10.1111/j.1476-5381.2009.00177.x] [PMID] [PMCID]
- [125] Amiel E, Ofir R, Dudai N, Soloway E, Rabinsky T, Rachmilevitch S. B-Caryophyllene, a compound isolated from the biblical balm of gilead (*commiphora gileadensis*), is a selective apoptosis inducer for tumor cell lines. *Evid Based Complement Alternat Med.* 2012; 2012:872394. [DOI:10.1155/2012/872394] [PMID] [PMCID]
- [126] Sain S, Naoghare PK, Devi SS, Daiwile A, Krishnamurthi K, Arrigo P, et al. Beta caryophyllene and caryophyllene oxide, isolated from aegle marmelos, as the potent anti-inflammatory agents against lymphoma and neuroblastoma cells. *Antiinflamm Antiallergy Agents Med Chem.* 2014; 13(1):45-55. [DOI:10.2174/18715230113129990016] [PMID]
- [127] Chen Z, Liu X, Gao W, Liu C-X. Studies on pharmacology, toxicology and pharmacokinetics of the chemical constituents in hemorrhheologic agent. *Asia J Pharmacokinet Pharmacodyn* 2008;8:15-30. https://www.researchgate.net/publication/238689755_Studies_on_pharmacology_agent
- [128] Seo EJ, Lee DU, Kwak JH, Lee SM, Kim YS, Jung YS. Antiplatelet effects of *Cyperus rotundus* and its component (+)-nootkatone. *J Ethnopharmacol.* 2011; 135(1):48-54. [DOI:10.1016/j.jep.2011.02.025] [PMID]
- [129] Choi HJ, Lee JH, Jung YS. (+)-Nootkatone inhibits tumor necrosis factor α /interferon γ -induced production of chemokines in HaCaT cells. *Biochem Biophys Res Commun.* 2014; 447(2):278-84. [DOI:10.1016/j.bbrc.2014.03.121] [PMID]
- [130] Ooi LS, Li Y, Kam SL, Wang H, Wong EY, Ooi VE. Antimicrobial activities of cinnamon oil and cinnamaldehyde from the Chinese medicinal herb *cinnamomum cassia blume*. *Am J Chin Med.* 2006; 34(3):511-22. [DOI:10.1142/S0192415X06004041] [PMID]
- [131] Subash Babu P, Prabuseenivasan S, Ignacimuthu S. Cinnamaldehyde--a potential antidiabetic agent. *Phytomedicine.* 2007; 14(1):15-22. [DOI:10.1016/j.phymed.2006.11.005] [PMID]
- [132] Mohamed SH, Hassan MA, Farag S, Mohamed GA. Phytochemical and biological studies of *Cyperus rotundus* L. growing in Egypt. *Bull Pharm Sci Cairo Univ* 2001; 39:195-203. https://www.researchgate.net/publication/284666083_Phytochemical_and_biological_investigations_of_Cyperus_rotundus_L
- [133] Sayed HM, Mohamed MH, Farag SF, Mohamed GA, Proksch P. A new steroid glycoside and furochromones

- from *Cyperus rotundus* L. *Nat Prod Res.* 2007; 21(4):343-50. [DOI:10.1080/14786410701193056] [PMID]
- [134] Soumaya KJ, Zied G, Nouha N, Mounira K, Kamel G, Genviève FD, et al. Evaluation of in vitro antioxidant and apoptotic activities of *Cyperus rotundus*. *Asian Pac J Trop Med.* 2014; 7(2):105-12. [DOI:10.1016/S1995-7645(14)60004-3] [PMID]
- [135] Singh AP, Sharma SK. A new pentacyclic triterpenoid with antimicrobial activity from the tubers of *Cyperus rotundus* Linn. *Hygeia JD Med.* 2015; 7(1):1-9. [DOI:10.15254/H.J.D.Med.7.2015.138]
- [136] Zhou, Z., Zhang, H. Phenolic and iridoid glycosides from the rhizomes of *Cyperus rotundus* L. *Med Chem Res.* 2013; 22(10):4830-5. [DOI:10.1007/s00044-013-0504-9]
- [137] Zhang T, Xu L, Xiao H, Zhou X, Mo S, Cai S, et al. A new iridoid glycoside from the rhizomes of *Cyperus rotundus*. *Bull Korean Chem Soc.* 2014; 35(7):2207-9. [DOI:10.5012/bkcs.2014.35.7.2207]
- [138] Jahan N, Khalil-Ur-Rahman AS, Asi MR. Phenolic acid and flavonol contents of gemmo-modified and native extracts of some indigenous medicinal plants. *Pak J Bot.* 2013; 45(3):1515-9.
- [139] Li Q, Yang W-y, Qu L-l, Qi H-Y, Huang Y, Zhang Z. Interaction of warfarin with human serum albumin and effect of ferulic acid on the binding. *J Spectrosc.* 2014; (ID834501):7. [DOI:10.1155/2014/834501] <https://www.hindawi.com/journals/jspec/2014/834501/>
- [140] Sayed HM, Mohamed MH, Farag SF, Mohamed GA, Omobuwajo OR, Proksch P. Fructose-amino acid conjugate and other constituents from *Cyperus rotundus* L. *Nat Prod Res.* 2008; 22(17):1487-97. [DOI:10.1080/14786410802038556] [PMID]
- [141] Zhou Z, Yin W, Zhang H, Feng Z, Xia J. A new iridoid glycoside and potential MRB inhibitory activity of isolated compounds from the rhizomes of *Cyperus rotundus* L. *Nat Prod Res.* 2013; 27(19):1732-6. [DOI:10.1080/14786419.2012.750318] [PMID]
- [142] Amesty Á, Burgueño-Tapia E, Joseph-Nathan P, Ravelo AG, Estévez-Braun A. Benzodihydrofurans from *Cyperus teneriffae*. *J Nat Prod.* 2011; 74(5):1061-5. [DOI:10.1021/np200020t] [PMID]
- [143] Zhou Z, Yin W. Two novel phenolic compounds from the rhizomes of *Cyperus rotundus* L. *Molecules.* 2012; 17(11):12636-41. [DOI:10.3390/molecules171112636] [PMID] [PMCID]
- [144] Aeganathan R, Rayar A, Ilayaraja S, Prabakaran K, Manivannan R. Anti-oxidant, antimicrobial evaluation and GC-MS analysis of *Cyperus rotundus* L. Rhizomes chloroform fraction. *Am J Ethno.* 2015 ;2(1):14-20. <https://www.imedpub.com/ethnomedicine/archive/ipaje-volume-2-issue-1-year-2015.htm>
- [145] Mohamed GA. Iridoids and other constituents from *Cyperus rotundus* L. Rhizomes. *Bulletin of Faculty of Pharmacy, Cairo University.* 2015; 53(1):5-9. [DOI:10.1016/j.bfopcu.2015.01.001]
- [146] Raut NA, Gaikwad NJ. Antidiabetic activity of hydro-ethanolic extract of *Cyperus rotundus* in alloxan induced diabetes in rats. *Fitoterapia.* 2006; 77(7-8):585-8. [DOI:10.1016/j.fitote.2006.09.006] [PMID]
- [147] Yazdanparast R, Ardestani A. [In vitro antioxidant and free radical scavenging activity of *Cyperus rotundus* (Persian)]. *J Med Food.* 2007; 10(4):667-74. [DOI:10.1089/jmf.2006.090] [PMID]
- [148] Ito T, Endo H, Shinohara H, Oyama M, Akao Y, Iinuma M. Occurrence of stilbene oligomers in *Cyperus* rhizomes. *Fitoterapia.* 2012; 83(8):1420-9. [DOI:10.1016/j.fitote.2012.08.005] [PMID]
- [149] Priya Rani M, Padmakumari KP. HPTLC and reverse phase HPLC methods for the simultaneous quantification and in vitro screening of antioxidant potential of isolated sesquiterpenoids from the rhizomes of *Cyperus rotundus*. *J Chromatogr B Analyt Technol Biomed Life Sci.* 2012; 904:22-8. [DOI:10.1016/j.jchromb.2012.05.042] [PMID]
- [150] Tran HH, Nguyen MC, Le HT, Nguyen TL, Pham TB, Chau VM, et al. Inhibitors of α -glucosidase and α -amylase from *Cyperus rotundus*. *Pharm Biol.* 2014; 52(1):74-7. [DOI:10.3109/13880209.2013.814692] [PMID]
- [151] Duarte MC, Figueira GM, Sartoratto A, Rehder VL, Delarmelina C. Anti-Candida activity of Brazilian medicinal plants. *J Ethnopharmacol.* 2005; 97(2):305-11. [DOI:10.1016/j.jep.2004.11.016] [PMID]
- [152] Soltan MM, Zaki AK. Antiviral screening of forty-two Egyptian medicinal plants. *J Ethnopharmacol.* 2009; 126(1):102-7. [DOI:10.1016/j.jep.2009.08.001] [PMID]
- [153] Eneh LK, Saijo H, Borg-Karlson AK, Lindh JM, Rajarao GK. Cedrol, a malaria mosquito oviposition attractant is produced by fungi isolated from rhizomes of the grass *Cyperus rotundus*. *Malar J.* 2016; 15:478. [DOI:10.1186/s12936-016-1536-7] [PMID] [PMCID]
- [154] Kaushik NK, Bagavan A, Rahuman AA, Mohanakrishnan D, Kamaraj C, Elango G, et al. Antiplasmodial potential of selected medicinal plants from eastern Ghats of South India. *Exp Parasitol.* 2013; 134(1):26-32. [DOI:10.1016/j.expara.2013.01.021] [PMID]
- [155] Kempraj V, Bhat SK. Ovicidal and larvicidal activities of *Cyperus giganteus* vahl and *Cyperus rotundus* Linn. Essential oils against aedes albopictus (Skuse). *Nat Prod Radianc.* 2008; (7)5:416-19. <http://nopr.niscair.res.in/bitstream/123456789/5712/1/%207%285%2920416-419.pdf>
- [156] Chang KS, Shin EH, Park C, Ahn YJ. Contact and fumigant toxicity of *Cyperus rotundus* steam distillate constituents and related compounds to insecticide-susceptible and -resistant *Blattella germanica*. *J Med Entomol.* 2012; 49(3):631-9. [DOI:10.1603/me11060] [PMID]
- [157] Puratchikody A, Devi CN, Nagalakshmi G. Wound healing activity of *Cyperus rotundus* linn. *Indian J Pharm Sci.* 2006; 68(1):97. [DOI:10.4103/0250-474X.22976]
- [158] Huang B, He D, Chen G, Ran X, Guo W, Kan X, et al. A-Cyperone inhibits LPS-induced inflammation in BV-2 cells through activation of Akt/Nrf2/HO-1 and suppression of the NF- κ B pathway. *Food Funct.* 2018; 9(5):2735-43. [DOI:10.1039/c8fo00057c] [PMID]
- [159] Ma Y, Zhao Y, Zhang R, Liang X, Yin Z, Geng Y, et al. A-Cyperone inhibits PMA-induced EPCR shedding through PKC pathway. *Biol Pharm Bull.* 2017; 40(10):1678-85. [DOI:10.1248/bpb.b17-00183] [PMID]

- [160] Seo YJ, Jeong M, Lee KT, Jang DS, Choi JH. Isocyperol, isolated from the rhizomes of *Cyperus rotundus*, inhibits LPS-induced inflammatory responses via suppression of the NF- κ B and STAT3 pathways and ROS stress in LPS-stimulated RAW 264.7 cells. *Int Immunopharmacol.* 2016; 38:61-9. [DOI:10.1016/j.intimp.2016.05.017] [PMID]
- [161] Soumaya KJ, Dhekra M, Fadwa C, Zied G, Ilef L, Kamel G, et al. Pharmacological, antioxidant, genotoxic studies and modulation of rat splenocyte functions by *Cyperus rotundus* extracts. *BMC Complement Altern Med.* 2013; 13:28. [DOI:10.1186/1472-6882-13-28][PMID] [PMCID]
- [162] Hussain M, Waqas HM, Hussain I, Majeed A, Raza SM, Janbaz KH. Pharmacological validation of the folkloric uses of *Cyperus rotundus* L. Indifferent ailments: An in vivo and in vitro research. *Pak J Pharm Sci.* 2018; 31(1):95-102. [PMID]
- [163] Yagi S, Babiker R, Tzanova T, Schohn H. Chemical composition, antiproliferative, antioxidant and antibacterial activities of essential oils from aromatic plants growing in Sudan. *Asian Pac J Trop Med.* 2016; 9(8):763-70. [DOI:10.1016/j.apjtm.2016.06.009] [PMID]
- [164] Sim Y, Choi JG, Gu PS, Ryu B, Kim JH, Kang I, et al. Identification of neuroactive constituents of the ethyl acetate fraction from cyperi rhizoma using bioactivity-guided fractionation. *Biomol Ther (Seoul).* 2016; 24(4):438-45. [DOI:10.4062/biomolther.2016.091] [PMID] [PMCID]
- [165] Peña-Fronteras JT, Villalobos MC, Baltazar AM, Merca FE, Ismail AM, Johnson DE. Adaptation to flooding in upland and lowland ecotypes of *Cyperus rotundus*, a troublesome sedge weed of rice: Tuber morphology and carbohydrate metabolism. *Ann Bot.* 2009; 103(2):295-302. [DOI:10.1093/aob/mcn085] [PMID] [PMCID]
- [166] Anyasi RO, Atagana HI. Profiling of plants at petroleum contaminated site for phytoremediation. *Int J Phytoremediation.* 2018; 20(4):352-61. [DOI:10.1080/15226514.2017.1393386] [PMID]
- [167] Basumatary B, Saikia R, Das HC, Bordoloi S. Field note: Phytoremediation of petroleum sludge contaminated field using sedge species, *Cyperus rotundus* (Linn) and *Cyperus brevifolius* (Rottb.) Hassk. *Int J Phytoremediation.* 2013; 15(9):877-88. [DOI:10.1080/15226514.2012.760520] [PMID]
- [168] Sundaramoorthy P, Chidambaram A, Ganesh KS, Unnikannan P, Baskaran L. Chromium stress in paddy: (I) nutrient status of paddy under chromium stress; (II) phytoremediation of chromium by aquatic and terrestrial weeds. *C R Biol.* 2010; 333(8):597-607. [DOI:10.1016/j.crvi.2010.03.002] [PMID]
- [169] Wang H, Liu Y, Wei S, Yan Z. Application of response surface methodology to optimise supercritical carbon dioxide extraction of essential oil from *Cyperus rotundus* Linn. *Food Chem.* 2012; 132(1):582-7. [DOI:10.1016/j.foodchem.2011.10.075] [PMID]

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