Larvicidal potential of *Cyathea* species against *Culex quinquefasciatus*

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Abstract

Resistance to insecticides has persuaded researchers to find new methods to control *Culex quinquefasciatus* proliferation. Plants may be a source of alternative agents for mosquito control due to ever-growing insecticide resistance in mosquito vectors and environmental imbalance caused by synthetic insecticides. The present study was intended to study the larvicidal activity of selected *Cyathea* species against the filarial vector *Culex quinquefasciatus*. Larvicidal potential of different extracts were evaluated and larval mortality were recorded. The larvae were more sensitive to ethanolic extracts of studied three *Cyathea* species when compared to other extracts. Acetone, chloroform and petroleum ether extracts were considered to be less effective. The LC50 values of different extracts ranged from 320.72 to 657.03 μg/ml. The results exhibited that the tested three *Cyathea* species showed concentration dependent potential larvicidal effects and also provide an indication of possible bioactive properties.

Keywords: Tree ferns, *culex quinquefasciatus*, larvicidal

Introduction

Vector-borne diseases are the major cause of morbidity in most of the tropical and subtropical countries. Mosquitoes are one of the important vectors responsible for the spread of several diseases viz., malaria, filariasis, dengue fever and Japanese encephalitis. *Culex* sp. is the most abundant mosquito species in urban areas. It is crucial to manage *Culex* population so that people can be protected from various mosquito borne diseases. These diseases can be controlled by targeting the causative parasites and pathogens. The chemical control was one of the most extensively used conventional methods for controlling mosquito borne diseases since chemical pesticides are relatively inexpensive and usually produces immediate control. Generally, the chemical control is carried out by the indoor residual spraying of insecticides such as temephos, insect growth regulators such as diflubenzuron and methoprene, which are still the most effective. Insecticide applications against the target species are facing a serious threat due to the development of resistance to chemical insecticides (1). Numerous studies have been carried out to identify safer mosquito control agents and to reduce environmental and human health concern (2, 3). An approach to obtain efficient, safe and selective insecticides is the study of natural products obtained from plants (4). In the world flora, next to angiosperms, pteridophytes occupy an important position in which many extant species were recorded (5). Pteridophytes are conspicuous and gorgeous elements of biodiversity which occurs in various kinds of habitats ranging from sea level to mountain top and tropical to subpolar regions (6). With reference to *Cyathea* species, Janakiraman and Johnson (7) studied the presence of phenolics, flavonoids, tannins, cardiac glycosides, terpenoids, steroids, saponins and alkaloids in *C. nilgirensis*, *C. gigantea* and *C. crinita*. Janakiraman and Johnson (8) determined the functional constituents of *C. nilgirensis*, *C. gigantea* and *C. crinita*. using FT-IR. Janakiraman and Johnson (9) studied the antioxidant potentials of *C. nilgirensis*, *C. gigantea* and *C. crinita*. Janakiraman and Johnson (10) revealed the phenolics profile of tree ferns using HPTLC analysis. Janakiraman and Johnson (11) studied the chemical constituents of tree ferns using GC-MS analysis. Janakiraman and Johnson (12) confirmed the cytotoxic activity of ethanolic extracts of selected *Cyathea* species against MCF 7 cell line cultures. Hence, the present study was undertaken to study the larvicidal potential of various extracts of *Cyathea nilgirensis* Hottum, *Cyathea gigantea* (Wall. ex. Hook.) Hottum and *Cyathea crinita* (Hook.) Copel.

Materials and methods

Collection of plant materials

Samples for the present study were collected from different parts of Tamil Nadu, South India.

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C. nilgirensis were harvested in and around Kakkachi stream (1,725 m), Tirunelveli hills (8° 44’ N and 77° 44’ E), C. gigantea from the road sides near Nadugani (2,637 m), Nilgiris hills (11° 24’ N and 76° 44’ E) and C. crinita from the Anglade Institute of Natural History, Shenbaganur, Kodaiakanal (2,195 m), Palni hills (10° 13’ N and 77° 32’ E), Western Ghats, South India. The specimens were identified based on the “Pteridophyte Flora of the Western Ghats, South India” by Manickam and Iruadayaraj (13). Herbarium specimens were deposited in the St. Xavier’s College Herbarium (XCH), Palayamkottai for further reference (C. nilgirensis - XCH 25423; C. gigantea - XCH 25422 and C. crinita - XCH 25424).

Preparation of extracts
The collected species of Cyathea were thoroughly washed with tap water followed by distilled water. They were blotted on the blotting paper and spread out at room temperature in shade to remove the excess water contents. The shade dried plant samples were ground to fine powder using mixer grinder. The powdered materials were stored in refrigerator for further use. Thirty gram of powdered sample was extracted (1:6 ratio) successively with 180 mL of petroleum ether, chloroform, acetone and ethanol using the Soxhlet extractor for 8-12 h at a temperature not exceeding the boiling point of the solvent. The extracts were concentrated in a vacuum at 40 °C using rotary evaporator.

Larvicidal activity
Culex quinquefasciatus (4th instar larvae) was collected from sewages of Tirunelveli district with the help of ‘O’ type brush. These larvae were brought to the laboratory and transferred to 18×13×4 cm size enamel trays containing 500 mL of water. It was maintained at 27 ± 2 °C, 75-85% RH and 14 h light and 10 h dark photoperiod cycles. Larvicidal activity of different extracts (petroleum ether, chloroform, acetone and ethanol) with varied concentrations of C. nilgirensis, C. gigantea and C. crinita was evaluated as per the standard method described by WHO (14). Batches of twenty 4th instar larvae of C. quinquefasciatus were collected separately and transferred to small disposable cups each containing 200 mL of water. The appropriate volume of dilution was added in the cups to obtain the desired target dosage (concentrations ranging from 100-500 µg/ml) starting with the lowest concentration. Five replicates were set up for each concentration and simultaneously a control was maintained. The larval mortality in both treated and control were recorded after 24 h. The standard larvicide Temephos (Abate) was used as positive control. The control mortality was corrected by Abbott’s formula (15).

Statistical analysis
The LC50 value was calculated by Probit analysis (16). The IC50 values of the crude extracts were found using MS-Excel 2007. Correlation analysis between LC50 and IC50 values were calculated using SPSS (Chicago, USA).

Results
The results of the larvicidal bioassay of crude petroleum ether, chloroform, acetone and ethanolic extracts of C. nilgirensis, C. gigantea and C. crinita against the fourth instar mosquito larvae C. quinquefasciatus were presented in Table 1. The larvae were more sensitive to ethanolic extracts of studied three Cyathea species when compared to other extracts. Acetone, chloroform and petroleum ether extracts were considered to be less effective. The LC50 values of different extracts ranged from 320.72 to 657.03 µg/ml. Among the three different species tested, the highest larval mortality was observed in ethanolic extracts of C. crinita with the LC50 value of 320.72 µg/ml followed by C. gigantea (361.07 µg/ml) and C. nilgirensis (373.99 µg/ml). The positive control Temephos showed 100% mortality rate at 0.025 mg/ml. The 95% confidence limits LC50 (LCL-UCL), LC90 and chi-square values were also calculated (Table 1). The Chi-square values for the studied extracts ranged from 0.35 to 7.37 for C. quinquefasciatus. Petroleum ether (0.35), chloroform (0.40) and ethanolic extracts (0.36) of C. crinita, chloroform (0.54) and acetone (0.38) extracts of C. gigantea and ethanolic extracts (0.69) of C. nilgirensis showed a strong positive correlation between the extracts and larval mortality (Table 1). The other tested extracts failed to show direct correlation between insect toxicity.

Discussion
The control of mosquito larvae by chemical substances is not safe at present because of environmental imbalance and insecticide resistance by vectors which leads to deleterious effects. The major drawback with the use of chemical insecticides is that they are non-selective and could be harmful to other organisms in the environment. Hence, an alternative mosquito control method is needed (17). The extracts which are obtained from plant parts have been used as conventional larvicide (18, 19). The observed results were also comparable with earlier reports. The fruit extract of Croton caudatus, flower extract of Tiliaacora acuminata (20), leaf extract of Typhonium trilobatum (21) and flower extract of Tagetes erecta (22) were found to cause larval mortality against C. quinquefasciatus. In the present study, the different extracts of C. nilgirensis, C. gigantea and C. crinita exhibited a dose dependent activity. The results observed were similar to previous studies which have also reported dose
Larvicidal potential of Cyathea

The tested three Cyathea species showed concentration dependent potential larvicidal effects. The results provide an indication of possible bioactive properties of the tested extracts. Further investigation is needed to find out the active compounds responsible for the larvicidal activity.

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Conflict of interest
The authors declare that they have no conflict of interest.

References

Table 1 Larvicidal activity of different Cyathea species against C. quinquefasciatus

| Species       | Extracts | $\text{LC}_{50}$ (µg/ml) | 95% Confidence Limits | $\text{IC}_{50}$ (µg/ml) | $\chi^2$ | $\Delta$ |
|---------------|----------|-------------------------|------------------------|--------------------------|---------|
|               |          | Lower | Upper | Lower | Upper |                             |         |
| C. nilgirensis | Pet. ether | 607.76 | 512.14 | 894.19 | 893.93 | 3.86 | 66.4 |
|               | Chloroform | 592.26 | 482.18 | 917.90 | 1078.70 | 2.73 | 52.3 |
|               | Acetone | 448.88 | 394.96 | 536.88 | 966.31 | 1.04 | 57.8 |
|               | Ethanol | 373.99 | 338.65 | 417.91 | 758.69 | 0.69 | 51.9 |
| C. gigantea   | Pet. ether | 639.92 | 561.76 | 784.12 | 1013.26 | 1.05 | 55.1 |
|               | Chloroform | 624.63 | 532.47 | 813.44 | 1151.44 | 0.54 | 51.4 |
|               | Acetone | 468.00 | 394.65 | 620.38 | 1175.06 | 0.38 | 141.6 |
|               | Ethanol | 361.07 | 317.84 | 416.12 | 848.87 | 2.29 | 64.8 |
| C. crinita    | Pet. ether | 675.03 | 556.55 | 868.54 | 1178.48 | 0.35 | 50.9 |
|               | Chloroform | 568.04 | 489.18 | 722.23 | 1103.12 | 0.40 | 51.9 |
|               | Acetone | 400.36 | 292.74 | 776.28 | 870.41 | 7.37 | 61.7 |
|               | Ethanol | 320.72 | 291.11 | 351.94 | 645.79 | 0.36 | 50.5 |
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